

*Assessment of NO<sub>x</sub> Emissions from Soil in California  
Cropping Systems*

FINAL REPORT

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# Determining NO<sub>x</sub> Emissions from Soil in California Cropping Systems to Improve Ozone Modeling

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## **Abstract**

Soils are a source of oxides of nitrogen ( $\text{NO}_x$  = nitric oxide and nitrogen dioxide), precursors for the production of ozone ( $\text{O}_3$ ), an air pollutant in the troposphere. Production of nitric oxide (NO) occurs through soil microbial processes using ammonium from nitrogen fertilizer and manure inputs or soil mineral nitrogen (N). Emissions of  $\text{NO}_x$  were measured in almond, alfalfa, tomato, wheat, and silage corn cropping systems during summer months to obtain estimates of  $\text{NO}_x$  emissions that could potentially be used in regional models predicting  $\text{O}_3$  in the San Joaquin Valley. The lowest average  $\text{NO}_x$  fluxes ( $<0.1 \text{ g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$ ) were measured at low soil moisture and in subsurface drip-irrigated tomato. The highest average emissions ( $0.5\text{--}2.8 \text{ g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$ ) occurred in high N input systems, such as silage corn. In alfalfa, almond, and furrow-irrigated tomato, average  $\text{NO}_x$  fluxes were intermediate ( $0.1\text{--}0.5 \text{ g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$ ). The  $\text{NO}_x$  emissions were related to N inputs, time since fertilizer applications, temperature, and soil moisture. Under field conditions  $\text{NO}_x$  fluxes increased 2.5-3.5-fold for each increase in soil temperature of  $10^\circ\text{C}$ . The  $\text{NO}_x$  emissions seem predictable in systems receiving N at recommended rates, ranging from  $0.02\text{--}2.5 \text{ g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$  in alfalfa, wheat, tomato, and almond, but in systems receiving large N inputs resulting in high concentrations of ammonium, episodes of very high  $\text{NO}_x$  emissions ( $>40 \text{ g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$ ) were measured. These high  $\text{NO}_x$  flux events are difficult to predict.

## **Executive Summary**

### *Background*

Soils are one of the sources of  $\text{NO}_x$  (nitric oxide and nitrogen dioxide), which is involved in reactions producing ozone ( $\text{O}_3$ ), a pollutant in the troposphere. In the San Joaquin Valley, ozone ( $\text{O}_3$ ) levels are often elevated during summer months at many locations. According to the California Emission Inventory Development and Reporting System (CEIDARS), in the San Joaquin Valley, the State's  $\text{O}_3$  standards were exceeded on more than 120 days per year during 2004-2007. Knowledge of the major sources of  $\text{NO}_x$  is essential to regionally predict  $\text{O}_3$  dynamics and evaluate the effectiveness of air quality management programs. To date, estimates of  $\text{NO}_x$  emitted from agricultural soil are not included in CEIDARS. Soil-borne production of  $\text{NO}_x$  occurs through soil microbial processes using ammonium from synthetic fertilizer and manure, and soil mineral nitrogen (N). The present study provides estimates of  $\text{NO}_x$  emissions during summer months from five cropping systems, comprising 17 different locations and management treatments.

### *Methods*

The  $\text{NO}_x$  emissions were measured in an almond orchard, and in alfalfa, tomato, wheat, and silage corn cropping systems following irrigation and nitrogen fertilization events mostly during June to September in 2011 and 2012. In the almond orchard, microjet sprinklers were used for irrigation and fertigation. Alfalfa and silage corn were flood irrigated. In the silage corn systems, synthetic fertilizer was applied before planting or as a side-dress and with most irrigations, liquid manure was mixed with the irrigation water. Tomato was either furrow-irrigated and most nitrogen fertilizer was applied as side-dress, or subsurface drip-irrigated and fertigated. Wheat, being a rainfed crop receiving N fertilizer in winter, was maturing at the time of the measurements. The  $\text{NO}_x$ -flux measurements were made by placing a chamber connected to a  $\text{NO}_x$ -analyzer on the soil surface for 3-5 minutes. The headspace air in the chamber was constantly circulated through the  $\text{NO}_x$  analyzer, and readings of the

concentration of  $\text{NO}_x$  were taken every 15 seconds. The flux was calculated on a per area basis by taking the rate of change of  $\text{NO}_x$  concentration, chamber volume and temperature into account. Soil moisture, soil ammonium and temperature were also measured to characterize how environmental conditions and management affected  $\text{NO}_x$  emissions.

### *Results*

The average hourly  $\text{NO}_x$ -fluxes were lowest ( $<0.1 \text{ g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$ ) in dry soil, such as maturing wheat and a tractor row in an almond orchard, and under subsurface drip irrigation in tomato. Intermediate fluxes ( $0.1\text{-}0.5 \text{ g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$ ) comparable to  $\text{NO}_x$  emissions reported in earlier studies were observed in almond, alfalfa, and in furrow-irrigated tomato fertilized at recommended N rates. The highest average hourly  $\text{NO}_x$  fluxes ( $0.4\text{-}2.8 \text{ g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$ ) took place in the systems receiving high N inputs, such as dairy silage corn and furrow-irrigated tomatoes fertilized at an excessive N rate. The emissions were related to N inputs leading to high soil ammonium concentrations. On some occasions the magnitude of  $\text{NO}_x$  emissions, which closely followed large N inputs of synthetic N fertilizer and/or liquid dairy manure, matched those of the highest fluxes ever measured. The  $\text{NO}_x$  fluxes decreased with time since N fertilization. Within a given day,  $\text{NO}_x$  fluxes increased 2.5-3.5-fold for each increase in soil temperature of  $10^\circ\text{C}$ . The  $\text{NO}_x$  fluxes were also dependent on soil water content with the highest fluxes occurring at intermediate soil moisture values (30-60% water-filled pore space) and lower fluxes at higher water content.

### *Conclusions*

The emissions at each location varied over time, depending on soil moisture, temperature, and time since N fertilization. The results suggest that  $\text{NO}_x$  emissions are related to ammonium availability and nitrification rates. Enhanced  $\text{NO}_x$  fluxes occurred under intermediate soil water contents (water-filled pore space 30-60%), whereas in relatively dry soils or at high water content,  $\text{NO}_x$ -fluxes were low. Field experiments showed that  $\text{NO}_x$  emissions increase on average 2.5- and 3.5-fold with each increase of  $10^\circ\text{C}$  in soil temperature at 1 and 5 cm depth, respectively. The study showed that  $\text{NO}_x$  fluxes are fairly predictable in cropping systems fertilized at recommended N rates ranging from  $0.02 - 2.5 \text{ g NO-N ha}^{-1} \text{ h}^{-1}$  in alfalfa, wheat, tomato, and almond. However, in the systems receiving high N inputs, such as silage corn, the emissions following N additions resulting in high soil ammonium concentrations can be enhanced by an order of magnitude, reaching hourly fluxes up to  $40 \text{ g NO-N ha}^{-1} \text{ h}^{-1}$  for several days. To regionally quantify  $\text{NO}_x$  emissions from agricultural land would require elaborate models that account for the mosaic of cropping systems and management events, such as N fertilization and irrigation, in individual fields.



## Introduction

Measurements of NO<sub>x</sub> (nitric oxide or NO and nitrogen dioxide or NO<sub>2</sub>) emissions from agricultural soil in the Central Valley, where ozone (O<sub>3</sub>) levels are often elevated during summer months at many locations, are needed as inputs in air quality models. Oxides of nitrogen (NO<sub>x</sub>) are required for O<sub>3</sub> formation. The build-up of O<sub>3</sub> depends on the ratio of volatile organic compounds (VOCs) to NO<sub>x</sub>, and the influence of these precursors on O<sub>3</sub> production varies temporally and spatially across the landscape (Blanchard and Fairley, 2001). When the ratio of VOCs to NO<sub>x</sub> is low, the availability of VOCs limits O<sub>3</sub> formation. However, when the ratio of VOCs to NO<sub>x</sub> is high, the availability of NO<sub>x</sub> controls O<sub>3</sub> formation, and under these conditions, reducing NO<sub>x</sub> will decrease O<sub>3</sub> production while reducing VOCs has little effect on O<sub>3</sub> formation. Therefore, quantifying all major NO<sub>x</sub> sources is essential to regionally predict the dynamics of O<sub>3</sub> in the troposphere and evaluate the effectiveness of air quality management programs.

About 16% of the world's annual NO<sub>x</sub> emissions originate from microbial activity in soils (Olivier *et al.*, 1998). Agricultural soils and associated fertilizer management are known to be sources of NO<sub>x</sub> (Williams *et al.*, 1995). However, only few data of NO<sub>x</sub> emissions from California agricultural soils have been reported (Matson and Firestone, 1995; Venterea and Rolston, 2000b; Lee *et al.*, 2009). Estimates of NO<sub>x</sub> emissions from biogenic (non-anthropogenic) and fertilizer applications are conspicuously absent in the California Emission Inventory Development and Reporting System (CEIDARS), and this lack of information restricts CARB's ability to develop accurate O<sub>3</sub> predictions through modeling.

The objectives of the present study were to determine NO<sub>x</sub> emissions in cropping systems typical for this region and to characterize NO<sub>x</sub> flux in response to various amounts of N fertilizer inputs under varying soil and air temperature conditions, with a focus on daytime emissions during summer months when O<sub>3</sub> concentrations are problematic in the San Joaquin Valley. This research benefits the staff of CARB and the San Joaquin Valley Air Pollution Control District by providing important data to improve modeling predictions of O<sub>3</sub> production.

## 2. Materials and Methods

### 2.1. Description of field sites by cropping system

This project assessed the NO<sub>x</sub> emissions in five different cropping systems including tomato, wheat, alfalfa, corn and almonds. Measurements of NO<sub>x</sub> fluxes in the different systems were carried out at sites selected for N<sub>2</sub>O emission monitoring in other projects commissioned by CARB ("Assessment of Baseline N<sub>2</sub>O Emissions in California Cropping Systems" and "Assessment of Baseline Nitrous Oxide Emissions in California's Dairy Systems, with Dr. Horwath as the PI). The different experimental sites were chosen so that a wide range of management strategies are represented such as different fertilizer inputs (inorganic N, manures), and irrigation systems (furrow, flood and sprinkler irrigation).

### 2.1.1. Almond

The NO<sub>x</sub> emissions in almond production systems were assessed in the Nickels Soil Laboratory in Colusa County, CA. Soil at this site is classified as a Fine-loamy, mixed, superactive, thermic Typic Haploxeralf with slightly acidic pH (Table 1). The trees were fertilized with 50 kg N ha<sup>-1</sup> as UAN32 four times during each summer 2011 and 2012. The NO<sub>x</sub> flux was measured following fertigation events in the tree rows where water and fertilizer solution were applied through microjet sprinklers and in the tractor rows, which were neither irrigated nor fertigated. The tractor row measurements thus served as experimental control. Sampling was carried out at 3 locations both in the tree and tractor rows.

**Table 1.** Soil (0-25 cm) characteristics at the almond site in Colusa County, CA (<http://casoilresource.lawr.ucdavis.edu/drupal/>).

Sand (%)	66.8
Silt (%)	19.2
Clay (%)	14
pH (H <sub>2</sub> O 1:1)	6.7
Bulk density (g cm <sup>-3</sup> )	1.62
Organic Matter (%)	0.75
Total N (g kg <sup>-1</sup> )	nd

### 2.1.2. Alfalfa

Two adjacent grower fields in the vicinity of Winters, CA, were used to measure NO<sub>x</sub> fluxes from alfalfa. The soil at this site is classified as a Myers clay, which is a fine, montmorillonitic, thermic Entic Chromoxerert (Table 2). One of the fields was a one year-old stand, the other a 5 year-old stand. Fields were flood irrigated approximately every 30 days. No N fertilizers were supplied. Alfalfa was harvested 6 times in 2011. Sampling was carried out on 8 dates, including immediately following a flood irrigation event, as well as on days when the fields were relatively dry. Measurements were made at six locations within each field.

**Table 2.** Soil characteristics (0-30 cm depth) of the alfalfa fields near Winters, CA.

Sand (%)	23
Silt (%)	43
Clay (%)	34
pH (H <sub>2</sub> O 1:1)	7.7
Bulk density 5-15 cm (g cm <sup>-3</sup> )	1.43
Total C (g kg <sup>-1</sup> )	12.58
Total N (g kg <sup>-1</sup> )	1.15

### 2.1.3. Tomato

Measurements in tomato systems were conducted at the UC Davis Russell Ranch Sustainable Agriculture research site. Soils at this site are classified as Yolo silt loam, a fine-silty, mixed, non-acid, thermic Typic Xerorthent and Rincon silty clay loam, a fine monmorillonitic, thermic Typic Haploxeralf (Table 3). The NO<sub>x</sub> fluxes were measured in the

tomato beds at several dates between May and August in furrow- and subsurface drip-irrigated systems. In the furrow-irrigated systems, the NO<sub>x</sub> fluxes were assessed at three levels of N fertilization, i.e. 0, 162, and 300 kg N ha<sup>-1</sup> in a conventional, winter-fallow tomato-wheat rotation, and additionally, a winter cover cropped (oats-vetch-bell beans mixture) system fertilized with 162 kg N ha<sup>-1</sup>. Fifty kg N ha<sup>-1</sup> were applied on April 12, 2011, as NPK-15-15-15 starter fertilizer (8.7% NH<sub>4</sub><sup>+</sup>, 6.3% NO<sub>3</sub><sup>-</sup>) in granular form banded at a depth of about 16 cm. The remainder of the N applications were applied as side dress N in the form of urea ammonium nitrate (UAN32), banded on May 13, three weeks after planting, at a depth of 17 cm. Furthermore, NO<sub>x</sub> fluxes were measured in subsurface drip-irrigated (SDI) tomato systems in two treatments. One was a winter-fallow and the other a cover-cropped system as above. Both systems were fertilized with a total of 179 kg N ha<sup>-1</sup>. The starter application was the same as in the furrow-irrigated systems, but the remainder of the N fertilizer was applied as UAN32 as fertigation of 22-33 kg N ha<sup>-1</sup> between May 19 and July 15, 2011. All treatments were replicated 3 times.

**Table 3.** Soil characteristics (0-30 cm depth) of the tomato cropping system at the UC Davis Russell Ranch Sustainable Agriculture facility.

Sand (%)	21.83
Silt (%)	47.00
Clay (%)	31.17
pH (H <sub>2</sub> O 1:1)	6.80
Bulk density beds 5-15 cm (Mg m <sup>-3</sup> )	1.37
furrows	1.52
Organic C (g kg <sup>-1</sup> )	10.30
Organic N (g kg <sup>-1</sup> )	1.00

#### 2.1.4 Wheat

Assessment of NO<sub>x</sub> flux from wheat systems was carried out in a grower field near Dixon, CA. The soil in this field is classified as a silty clay loam thermic Typic Chromoxerert, its main physical characteristics are summarized in Table 4. The NO<sub>x</sub> flux was measured in beds and furrows in three treatments at the end of May: 0, 210 kg N ha<sup>-1</sup> applied either as ammonium sulfate and urea or as anhydrous ammonia and urea (112 kg N ha<sup>-1</sup> as starter in early November, and 98 kg N ha<sup>-1</sup> as aerial application in February). In addition to winter rainfall, the field received irrigation from April 16-22. The NO<sub>x</sub> flux was also measured in a wheat field at the Russell Ranch Sustainable Agriculture research site (see Table 1 for main physicochemical characteristics). Flux measurements from the Dixon field were collected in 2011 while the measurements at the Russell Ranch were made in 2012. At the Russell Ranch, 112 kg N ha<sup>-1</sup> was applied in the form of urea as starter, and 80 kg N ha<sup>-1</sup> was added as foliar N application in early March. In both fields NO<sub>x</sub> flux was assessed in beds and furrows, and values were weighted according to area (70% bed, 30% furrow) to calculate average field emissions. Two replications were used per treatment (n=2).

**Table 4.** Main soil characteristics (0-30 cm depth) at the wheat field located near Dixon, CA.

Sand (%)	21.3
Silt (%)	43.7
Clay (%)	35
pH (H <sub>2</sub> O 1:1)	7.4
Bulk density 5-15 cm (Mg m <sup>-3</sup> )	1.29
Total C (g kg <sup>-1</sup> )	14.9
Total N (g kg <sup>-1</sup> )	1.3

#### 2.1.5. Dairy silage corn

Assessment of NO<sub>x</sub> flux in corn systems was carried out in three forage production systems surrounding dairy farms, located in the counties of Stanislaus (Farms A and B) and Sacramento (Farm C). It is characteristic of the dairy farms in the Central Valley of California to use the farmland surrounding the facilities to produce silage corn and other forage crops which are in part fertilized with the manure generated at the dairy farms. Forage cropland land typically receives high annual inputs of nitrogen (N) compared to other cropping systems. According to our previous research, the N inputs into these silage corn/winter forage cropping systems range from 500 to 1200 kg N ha<sup>-1</sup> yr<sup>-1</sup>, versus 350 to 600 kg N ha<sup>-1</sup> yr<sup>-1</sup> removed in the harvested crop (Geisseler *et al.*, 2012). Manure generated at the farms is either collected as solids from the stables and applied to the fields or flushed with water and stored in anaerobic ponds. The liquid effluent is processed before storage in the lagoons to separate particles larger than a few millimeters from the liquid components. Manure is generally applied as liquid (“lagoon water”) and mixed with the irrigation water, although fields may also receive the different forms of solid manure. Liquid manure with high concentrations of NH<sub>4</sub><sup>+</sup> is diluted with irrigation water. In addition, inorganic N fertilizers are added. Irrigation is carried out through flooding of the fields. Monitoring of the NO<sub>x</sub> fluxes was carried out before and after the irrigation events at four locations within each field (n=4).

**Table 5.** Soil characteristics (0-30 cm depth) of the silage corn systems.

	Farm A		Farm B		Farm C
	Field 1	Field 2	Field 1	Field 2	
Sand (%)	78	70	84	84	31
Silt (%)	16	23	12	10	28
Clay (%)	7	7	4	6	41
pH (H <sub>2</sub> O 1:1)	6.7	7.2	6.8	7.3	7.47
Bulk density 5-15 cm (Mg m <sup>-3</sup> )	1.67	1.43	1.37	1.44	1.51
Total C (g kg <sup>-1</sup> )	10.4	12.5	11.8	6.8	12.4
Total N (g kg <sup>-1</sup> )	1	1.2	1.1	0.6	1.3

**Table 6.** Dairy silage corn nitrogen inputs and application method.

Year	Dates		N inputs					Total available N per fertigation event (kg N ha <sup>-1</sup> )
	Planting	Harvest	Synthetic Fertilizer-N		Manure- N			
			Rate (kg N ha <sup>-1</sup> )	Application method	Soluble (kg N ha <sup>-1</sup> )	Solids (kg N ha <sup>-1</sup> )	Application method	
<i>Farm A</i>								
2011	15 April	22 August	298	Irrigation water	198	69	Irrigation water	20-40
2012	6 May	24 August	182	Irrigation water	172	114	Irrigation water	20-40
<i>Farm B</i>								
2011	15 May	31 August.	104	Injected	245	713	Solid & irrigation water	20-50
2012	13 May	3 September	118	Injected	268	72	Irrigation water	20-50
<i>Farm C</i>								
2011	20 June	14 October	224	Injected	159	35	Irrigation water	159
2012	18 June	21 October	224	Injected	460	100	Irrigation water	89-115

At Farm A, two fields characterized as a coarse-loamy, mixed, active, thermic Typic Haploxeralf with neutral to slightly acidic pH, a total soil C content in the topsoil of 10.4 and 12.5 g kg<sup>-1</sup> soil, and a high sand content (70-78%)(Table 5). Fertilizer inputs consisted mainly of lagoon water and inorganic N fertilizer (UAN32) applied via the irrigation water (20-40 kg available N ha<sup>-1</sup> per irrigation event). The total N inputs were categorized as synthetic mineral N, solids (organic N, which has to undergo mineralization before becoming available to plants and microbes), or soluble organic N, which is readily available for plant or microbial uptake (Table 6).

The soil in Farm B was classified as a mixed, thermic Typic Xeropsamment. It was characterized by a high sand content (84%), a neutral to acidic pH, and a total C content in the topsoil of 11.8 g kg<sup>-1</sup> soil. Solid corral manure, partially composted, and the solid fraction, so called 'separator manure', remaining after mechanical separation of the liquid manure were incorporated into the soil in spring 2011 after disking and two weeks before corn planting. Synthetic N fertilizer as UAN was applied in early June at the rate of 104 and 117 kg N ha<sup>-1</sup> in 2011 and 2012, respectively. In addition, lagoon water was mixed into the irrigation water (approx. mixing rate 3:1 fresh water: lagoon water) resulting in N applications of 20-50 kg available N ha<sup>-1</sup> per irrigation event.

Soil at Farm C was classified as a fine, mixed, superactive, thermic Abruptic Durixeralf. It was characterized by a neutral pH (7.47), a total soil C in the topsoil of 12.4 g kg<sup>-1</sup>, and a lower sand and higher clay content than on Farms A and B. The main N inputs to the corn crop consisted on 227 kg ha<sup>-1</sup> of anhydrous ammonia injected in the soil one week before planting, and the application of lagoon water through irrigation (mixing rate 1:1 water: lagoon water), on August 29, 2011 (159 kg available N ha<sup>-1</sup>) and September 12 and 27, 2012 (115 and 89 kg available N ha<sup>-1</sup>, respectively).

## 2.2. *NO<sub>x</sub> flux measurements*

The NO<sub>x</sub> flux measurements took place during the summer months (2011 and 2012) when O<sub>3</sub> reaches critical threshold values, and measurements focused on soil fertilization events (if applicable, e.g. alfalfa is typically not fertilized) and varied soil moisture conditions. Following N fertilization events in the almond orchard, NO<sub>x</sub> fluxes were measured every 2-3 days until the fluxes subsided to the magnitude measured prior to these events, normally after 7-10 days. In addition, the fluxes were measured a few times in between the fertilization events to confirm background values. In alfalfa, we measured the NO<sub>x</sub> fluxes every other day during two irrigation events, as well as twice in between irrigations to obtain background values. In tomato, we measured the NO<sub>x</sub> fluxes every 2-3 days following two irrigation events. In the silage corn systems, the NO<sub>x</sub> fluxes were measured every other day during 5 irrigation events, which coincided with fertilizer applications since on two farms, lagoon water and/or synthetic N fertilizer is mixed with the irrigation water. On Farm C, the NO<sub>x</sub> flux was measured almost every day during three periods, twice following synthetic N fertilizer applications and once following lagoon water application. In wheat, we measured NO<sub>x</sub> flux twice.

Nitric oxide flux was measured in the field by using a dynamic chamber method. Either rectangular or circular chambers were used depending on the cropping system. Rectangular stainless steel chamber bases were 50 x 30 cm and 8 cm deep with a 2 cm-wide horizontal flange at the top end. Chamber bases were inserted in the soil so that the flanges rested in the

soil surface and were left in place unless field operations required their temporary removal. Thin-wall stainless steel (20 gauge) chamber tops (50 x 30 x 10 cm), with flanges facing down and lined with a rubber gasket, were placed onto the bases and secured with metal clamps. In the alfalfa systems, smaller rectangular chambers and bases were used (13.5 x 15 x 10 cm). Round, 20 cm diameter 10 cm tall polyvinyl chloride (PVC) chambers were used in wheat and almond systems. Chambers were placed on PVC rings buried 6-8 cm deep into the soil and sealed with a rubber gasket.

Both rectangular and round chambers were equipped with inlet and outlet ports connected to a chemoluminescence  $\text{NO}_x$  analyzer (LMA-3, Scintrex/Unisearch Associates, Concord, ON, Canada) via Teflon<sup>TM</sup> (PTFE) tubing covered by a layer of opaque material to prevent exposure of the air stream to sunlight. Chamber air was continuously circulated by a pump in the  $\text{NO}_x$  analyzer, flowing through a chromium tri-oxide ( $\text{CrO}_3$ ) column converting NO to  $\text{NO}_2$  and then the luminol chemiluminescence detector.  $\text{NO}_x$  concentrations were recorded every 15 seconds during 5 minutes right after sealing the chamber. The instrument was calibrated in the laboratory before every field measurement event by mixing NO-free air scrubbed of  $\text{NO}_x$  by permanganate-coated porous silica and a  $\text{NO}_x$  gas standard (Scott Marrin, Inc., Riverside, CA) in varied proportions and at known flow rates (Manostat). Upon return to the laboratory after field measurements, the calibration was checked. On most days, values stayed within 10% of the calibration values. On some days, when re-check values diverged from the calibrated ones obtained in the morning, the results were adjusted to a new calibration curve, representing a mean value of before- and after-field measurements. On a few days, field measurement values were discarded due to instrument problems.

The  $\text{NO}_x$  flux was calculated from the rate of change in chamber concentration, chamber volume, and soil surface area. Chamber gas concentrations determined by the  $\text{NO}_x$ -analyzer (volumetric parts per billion) were converted to mass per volume units assuming ideal gas relations using chamber air temperature values, which were measured by a thermocouple thermometer during each sampling event. The hourly flux was derived from the flux measured during 2-5 minutes at each chamber location. Typically fluxes at different locations within the same field were measured during the course of 1 hour or more, so we report mean hourly fluxes. The  $\text{NO}_x$  fluxes were converted to  $\text{g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$  to illustrate the relationship between  $\text{NO}_x$  flux and N inputs, which are usually reported as  $\text{kg ha}^{-1}$ , and because individual chambers (sampling replicates) in a given system were dispersed over large areas encompassing one to several hectares.

### 2.3. *Effects of environmental variables on $\text{NO}_x$ flux*

#### 2.3.1. *Soil chemical and physical analyses*

In order to gain a better understanding of the conditions affecting  $\text{NO}_x$  emissions, key soil and environmental variables were recorded in addition to the main management practices and N inputs at each site. Inorganic N ( $\text{NO}_3^-$  and  $\text{NH}_4^+$ ) content was determined in the 0-15 cm soil layer. Soil samples were collected close to the gas chamber bases by using a 1.83-cm steel corer. Soil samples were immediately extracted with 1 M potassium chloride (KCl) at a 1:5 ratio (soil: extracting solution) and analyzed within one day for nitrite ( $\text{NO}_2^-$ ). Extracts were also analyzed colorimetrically for ammonium ( $\text{NH}_4^+$ ) and nitrate ( $\text{NO}_3^-$ ) by a Shimadzu spectrophotometer (Model UV-Mini 1240). For determining  $\text{NH}_4^+$ , the phenate (indophenol blue) method was employed (Forster, 1995). Nitrate in the extracts was reduced to nitrite ( $\text{NO}_2^-$ ) with vanadium chloride, and the  $\text{NO}_2^-$  was analyzed by diazotizing with sulfanilamide followed by coupling with N-(1-naphthyl) ethylenediamine-dihydrochloride (Doane and

Horwáth, 2003). The pH was determined in the supernatant of soil slurries with H<sub>2</sub>O by a pH meter (Model 220, Denver Instrument Co., Arvada, CO).

Gravimetric soil moisture was calculated from field-moist and oven-dry (105°C, 24h) mass of soil collected in the field. Soil texture was determined by a modified pipet method (USDA, 1992). The bulk density was measured twice per growing and rainy season by collecting 10 cm dia. x 6 cm long cores in the 5-15 cm layer of soil, followed by drying the cores to 105°C. Water filled pore space (WFPS) was calculated from the gravimetric moisture (w) and measured bulk density values in the 5-15 cm layer as follows:

$$\% \text{WFPS} = (w * \text{bulk density}) / [1 - (\text{bulk density}/2.65)] * 100\%$$

where w = gravimetric water content.

### 2.3.2. Air and soil temperature

Air and soil temperature at 1 and 5 cm depths were routinely recorded simultaneously with NO<sub>x</sub> flux determinations at each individual sampling location. In addition, the relationship between soil temperature and NO<sub>x</sub> flux was explored at several field sites by taking measurements of NO<sub>x</sub> flux and temperature over the entire course of some days. One of the sites was a sandy loam soil planted to corn and fertilized with 180 kg N ha<sup>-1</sup> in the form of UAN32 near the UC Davis campus. The other sites were the field locations on Farms B and C. Q10 values were calculated as

$$Q10 = (\text{NO}_x\text{-flux}_2 / \text{NO}_x\text{-flux}_1)^{10 / (T_2 - T_1)}$$

where T<sub>1</sub> is the temperature in (°C) at 1 or 5 cm depth during NO<sub>x</sub>-flux measurement 1 and T<sub>2</sub> the corresponding temperature measurement at NO<sub>x</sub>-flux measurement 2.

More precisely, soil temperature at 1 and 5 cm depth, and ambient air temperature were recorded simultaneously with NO<sub>x</sub> flux determinations at each individual sampling location.

## Results

A summary of average hourly NO<sub>x</sub> fluxes among all the measurements taken in the different systems is shown in Table 7. The fluxes and results of the ancillary data are explained in detail below and in the accompanying figures.

### 3.1. Almonds

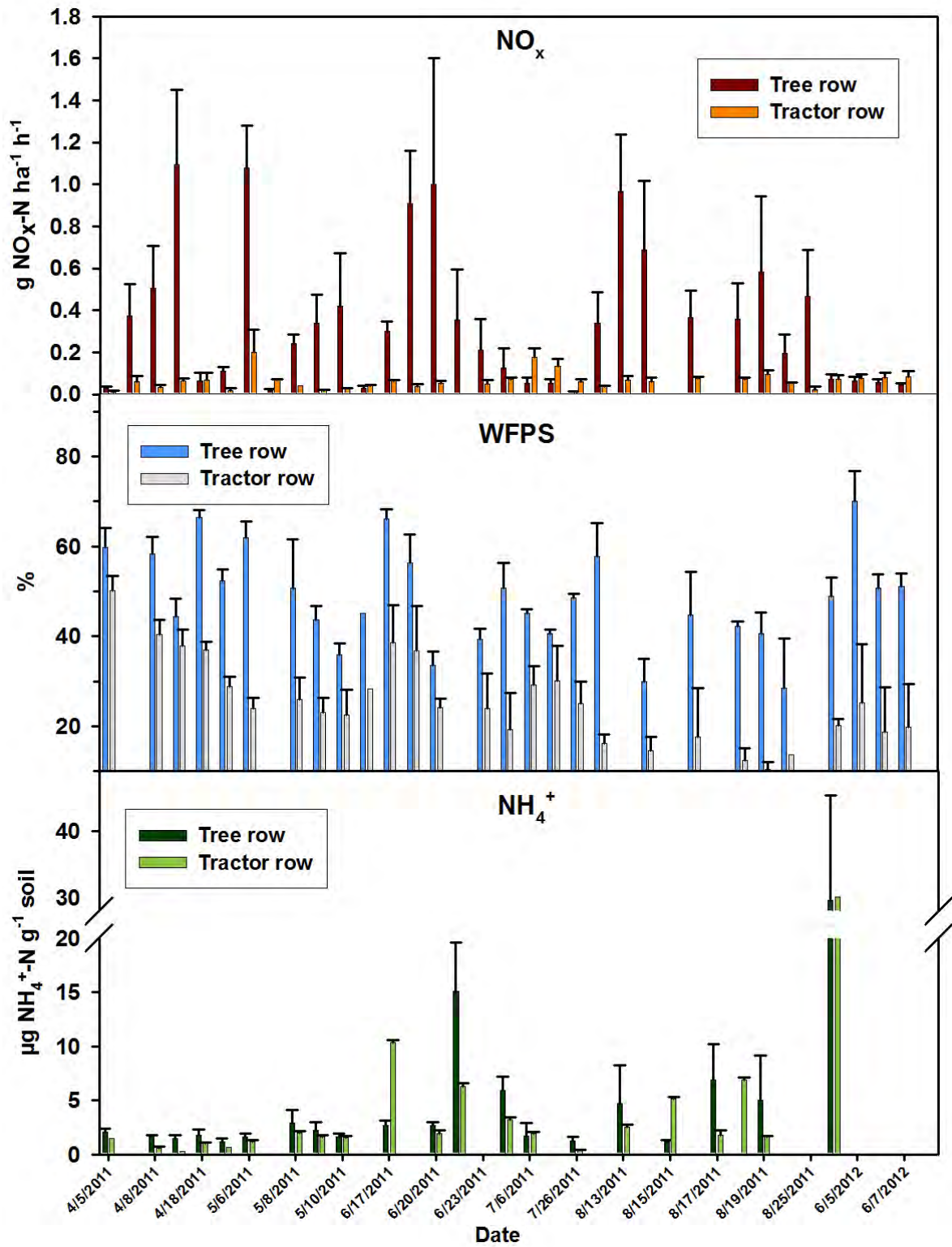
The NO<sub>x</sub>-flux was higher in the irrigated, fertigated tree than the tractor rows, which remained dry all summer. Following the monthly N fertilizer applications, the NO<sub>x</sub> flux reached about 1 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> and then gradually declined over the course of about 10 d to <0.1 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> (Figure 1). Baseline NO<sub>x</sub> emissions in tractor rows were almost always <0.1 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup>. In the tree rows, there was a weak correlation of NO<sub>x</sub>-flux with nitrite (NO<sub>2</sub><sup>-</sup>) (Figure 2). The soil NH<sub>4</sub><sup>+</sup> concentrations did not fluctuate much with the N fertilizer application and were similar in magnitude in the tree and tractor rows. The tractor rows had lower soil moisture content and higher soil temperatures than the tree rows.

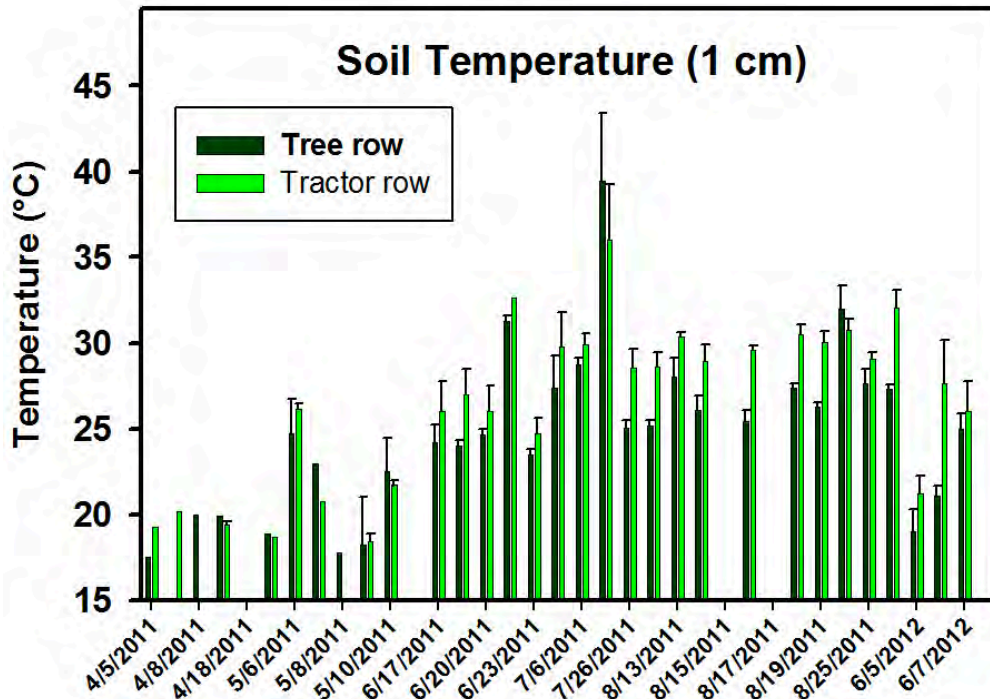


**Table 7.** Summary of mean hourly NO<sub>x</sub> fluxes and standard errors (SE) among means per location in different cropping systems.

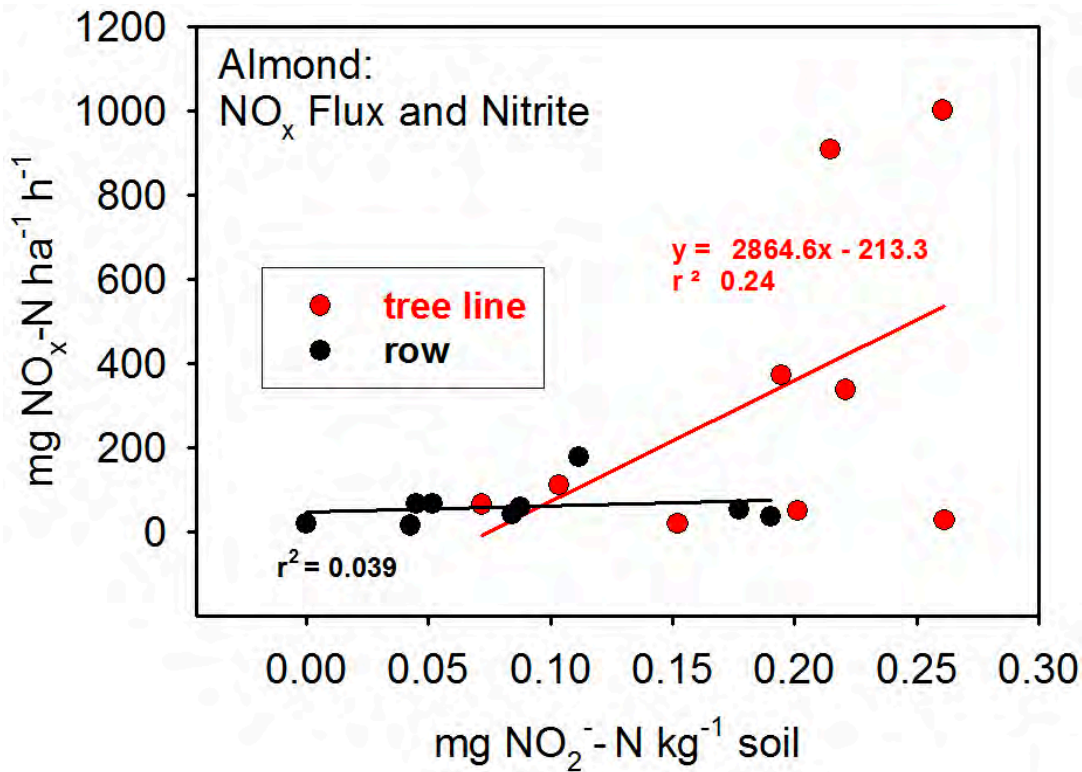
	# of Measurements	Chamber locations	Average flux		Average flux	
			g NO <sub>x</sub> -N ha <sup>-1</sup> h <sup>-1</sup>		g N <sub>2</sub> O-N ha <sup>-1</sup> h <sup>-1</sup>	
				SE		SE
Wheat (210 <sup>*</sup> , 192 <sup>**</sup> kg N ha <sup>-1</sup> )	9	3	0.04	0.01	0.09	0.02
Alfalfa, 5 year old stand	42	6	0.19	0.02	3.10	1.2
1 year old stand	48	6	0.54	0.31	1.67	0.46
Almond, tractor row	93	3	0.06	0.01	nd	
Tree row (200 kg N ha <sup>-1</sup> )	96	3	0.35	0.05	nd	
Tomato, SDI winter-fallow (179 kg N ha <sup>-1</sup> )	57	3	0.07	0.01	0.11	0.02
SDI, winter cc (179 kg N ha <sup>-1</sup> )	57	3	0.18	0.02	0.15	0.03
FI, zero N applied	60	3	0.10	0.03	0.11	0.03
FI, standard N rate (162 kg ha <sup>-1</sup> )	72	3	0.22	0.05	0.43	0.08
FI, standard N rate, cc	66	3	0.32	0.15	0.88	0.22
FI, 300 kg N ha <sup>-1</sup> applied	75	3	2.79	0.64	1.22	0.19
Silage corn, Farm A, Field 1 (565 <sup>*</sup> , 468 <sup>**</sup> kg N ha <sup>-1</sup> )	72	4	0.75	0.04	4.13	0.92
Farm A, Field 2 (565 <sup>*</sup> , 468 <sup>**</sup> kg N ha <sup>-1</sup> )	56	4	0.39	0.06	3.20	1.45
Farm B, Field 1 (1062 <sup>*</sup> , 458 <sup>**</sup> kg N ha <sup>-1</sup> )	88	4	2.03	0.28	4.8	0.96
Farm B, Field 2 (1062 <sup>*</sup> , 458 <sup>**</sup> kg N ha <sup>-1</sup> )	44	4	1.98	0.49	1.72	0.28
Farm C (421 <sup>*</sup> , 784 <sup>**</sup> kg N ha <sup>-1</sup> )	324	4	1.61	0.51	6.75	0.90

SDI = Subsurface drip-irrigation; FI = Furrow irrigation; cc = cover crop; <sup>\*</sup> 2011, <sup>\*\*</sup> 2012





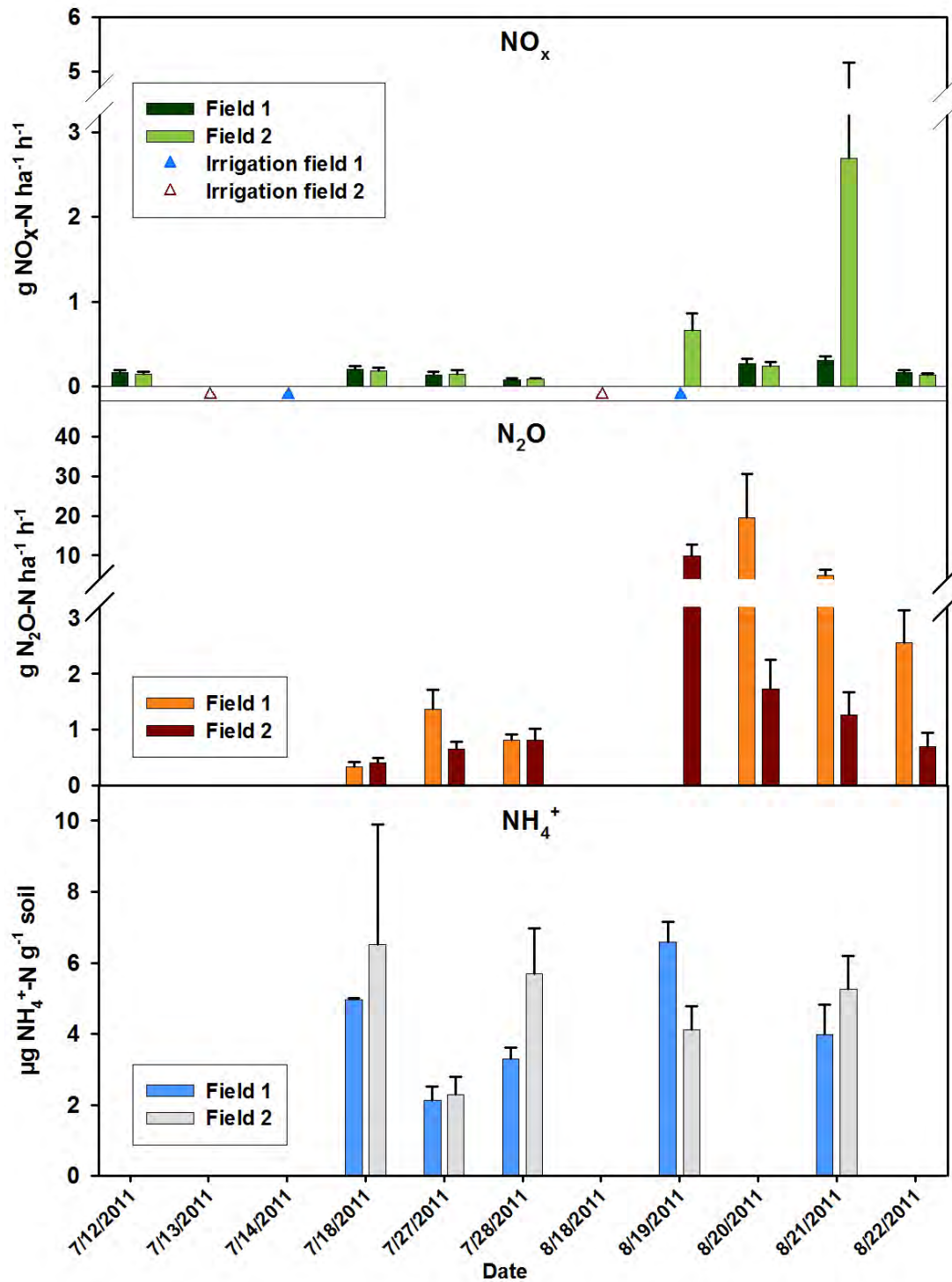
**Figure 1.** NO<sub>x</sub> flux, water-filled pores space (WFPS), ammonium (NH<sub>4</sub><sup>+</sup>) concentrations, and soil temperature at 1 cm depth in the almond orchard. Standard errors shown as line bars. n=3.

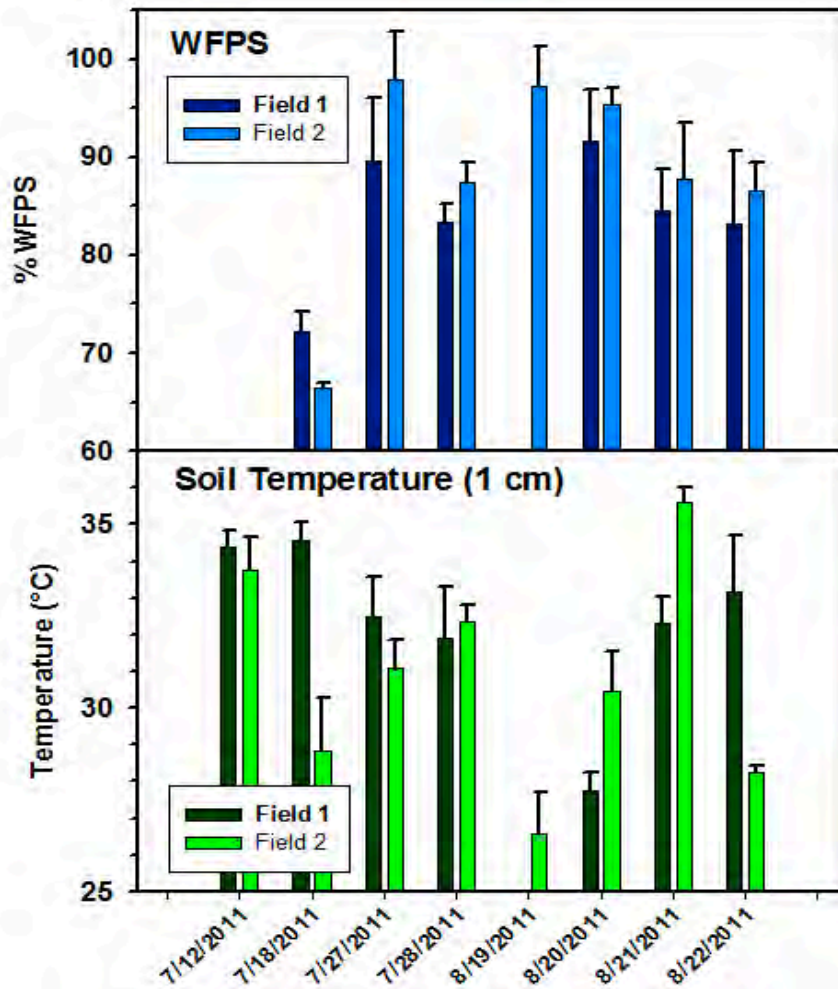


**Figure 2.** NO<sub>x</sub> flux vs. nitrite (NO<sub>2</sub><sup>-</sup>) concentrations in the soil in the almond orchard.

### 3.2. Alfalfa

In flood-irrigated alfalfa, the  $\text{NO}_x$  fluxes were generally  $<0.3 \text{ g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$  except for one event following an irrigation application when the  $\text{NO}_x$  flux reached almost  $3 \text{ g NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$  (Figure 3). This enhanced  $\text{NO}_x$  flux occurred in the 1 year-old field two days after an irrigation event. At this event, the  $\text{NO}_x$  was greater than the  $\text{N}_2\text{O}$  flux, but in general,  $\text{N}_2\text{O}$  was greater than  $\text{NO}_x$  flux by an order of magnitude.





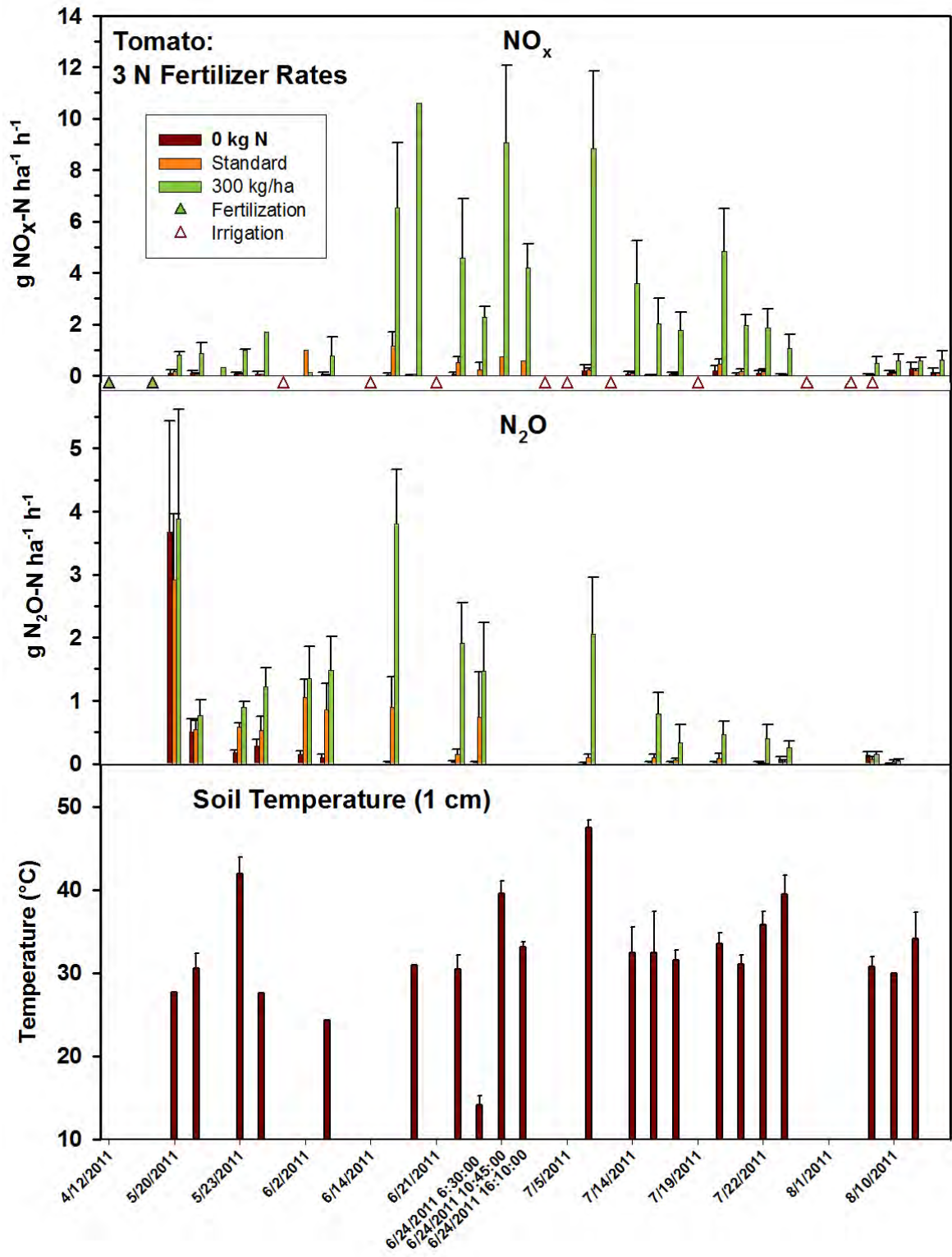
**Figure 3.** NO<sub>x</sub> and nitrous oxide (N<sub>2</sub>O) flux, soil ammonium (NH<sub>4</sub><sup>+</sup>) concentrations, water-filled pores space (WFPS), and soil temperature at 1 cm depth in the alfalfa fields. Standard errors are shown as line bars. n=6. Triangles along the x-axis indicate irrigation events for Field 1 (blue) and Field 2 (white).

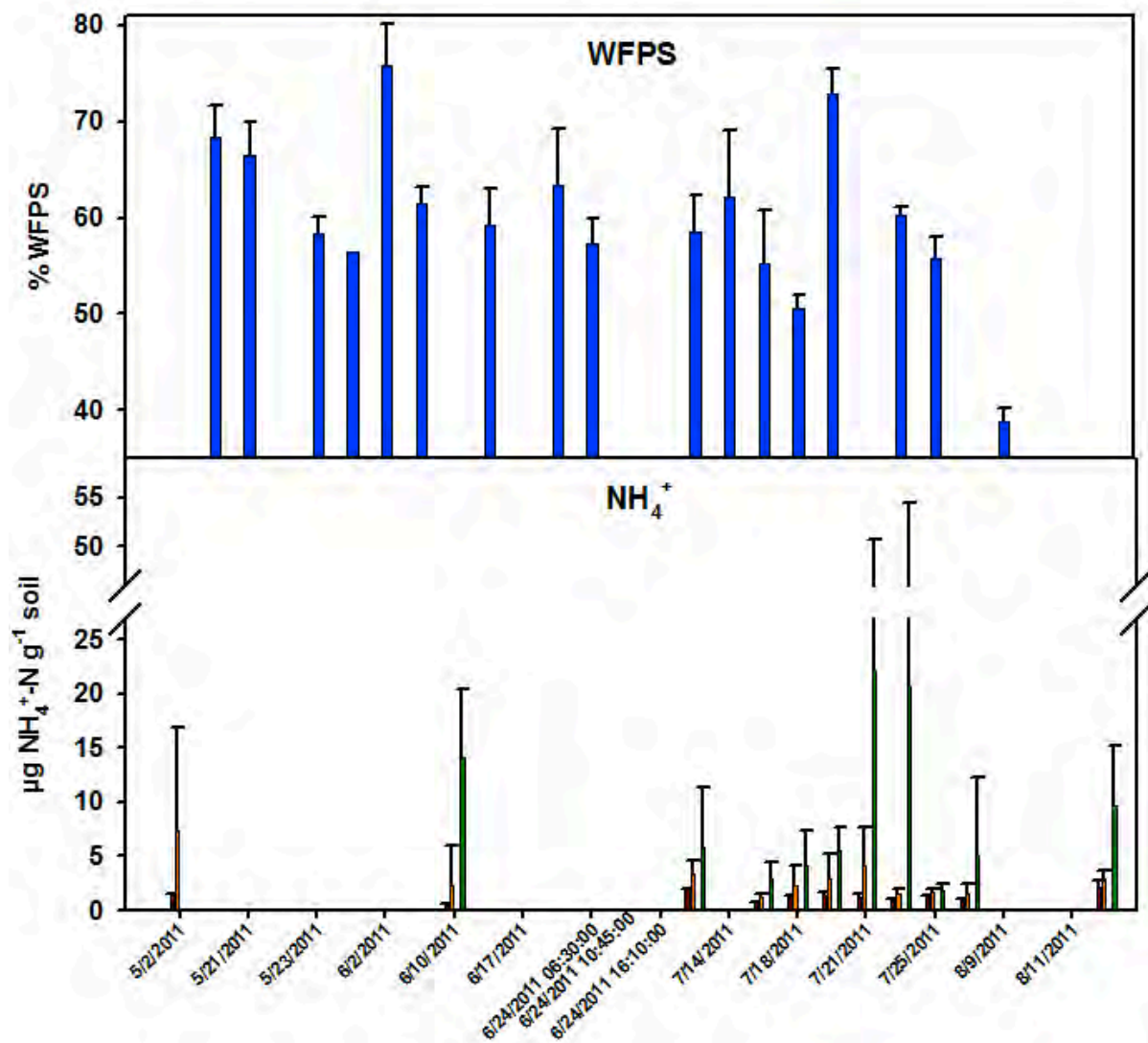
In both fields soil NH<sub>4</sub><sup>+</sup> concentrations were 2-6 μg NH<sub>4</sub><sup>+</sup>-N g<sup>-1</sup>. The soils almost reached saturation (100% WFPS) after irrigations and declined to 60-70% WFPS in between irrigations. Soil temperatures were lowest immediately following an irrigation and increased with drainage.

### 3.3. *Tomato*

The  $\text{NO}_x$  flux ranged from 1-10  $\text{g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$  in the tomato system fertilized with 300  $\text{kg N ha}^{-1}$  (high N treatment),  $<1 \text{ g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$  in the system fertilized at the rate of 162  $\text{kg N ha}^{-1}$  (standard N treatment), and  $<0.1 \text{ g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$  in the plots that did not receive any N fertilizer (zero N treatment) in that season (Figure 4). Most measurements were taken at temperatures (soil temperature at 1 cm depth) around 30°C. On June 24, 2011, at 6:30 am, the soil temperature was 14.1°C, increasing to 39.6°C by 10:45 am, and declining to 33.0°C by 4 pm. Concurrently,  $\text{NO}_x$  flux in the high N treatment increased from 2.3 to 9.1, and decreased to 4.2  $\text{g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$ . In the standard treatment,  $\text{NO}_x$  flux increased from 0.2 to 0.8, and declined to 0.6  $\text{g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$  during that day. On the days both  $\text{NO}_x$  and  $\text{N}_2\text{O}$  flux were measured, the amount of N emitted as  $\text{NO}_x$  was on average about three times greater than the amount of  $\text{N}_2\text{O-N}$  in the high N treatment and about equal to that in the standard N treatment. The soil water content in the furrow-irrigated tomatoes was mostly between 55 and 70%. Soil  $\text{NH}_4^+$  concentrations were 5-20  $\mu\text{g NH}_4^+\text{-N g}^{-1}$  soil in the high N treatment, 1-3  $\mu\text{g NH}_4^+\text{-N g}^{-1}$  soil in the standard N treatment, and  $<1.5 \mu\text{g NH}_4^+\text{-N g}^{-1}$  soil in the zero N treatment.

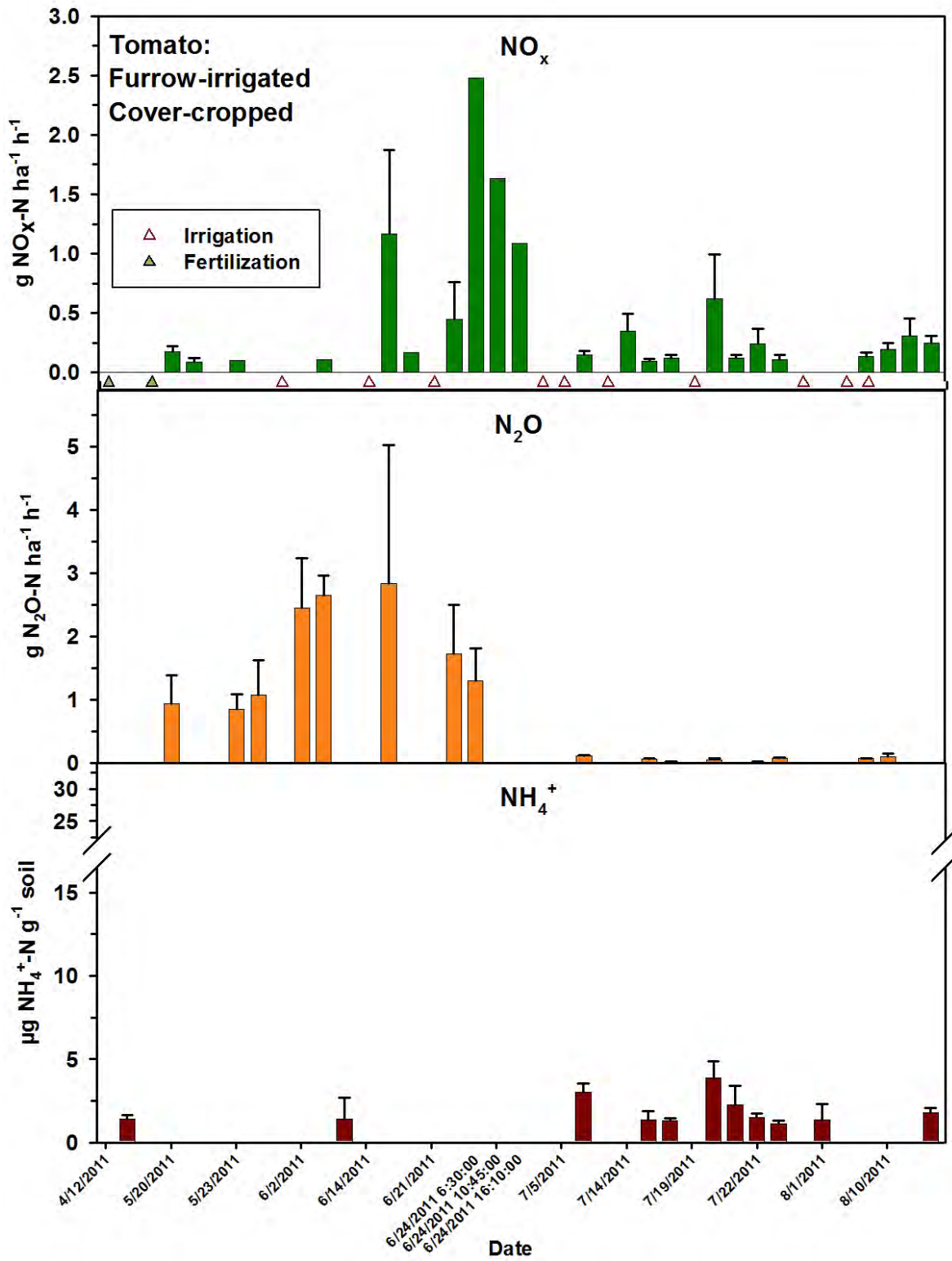
In the winter cover-cropped tomato system, the  $\text{NO}_x$  fluxes reached as much as 2.5  $\text{g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$  on some days, but during the remaining period were mostly  $<0.5 \text{ g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$  (Figure 5). The corresponding  $\text{N}_2\text{O-N}$  fluxes were similar in magnitude as the  $\text{NO}_x$  fluxes. With SDI, the  $\text{NO}_x$  fluxes ranged from 0.01-0.3  $\text{g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$  (Figure 6). On most days when the fluxes were measured, the  $\text{NO}_x\text{-N}$  emissions were greater in the system a cover crop containing legumes had been grown the previous winter, whereas soil  $\text{NH}_4^+$  concentrations were similar between the two SDI systems. The  $\text{N}_2\text{O}$  fluxes ranged from 0.05-0.5  $\text{g N}_2\text{O-N ha}^{-1} \text{h}^{-1}$ . The WFPS ranged from about 35-50%.

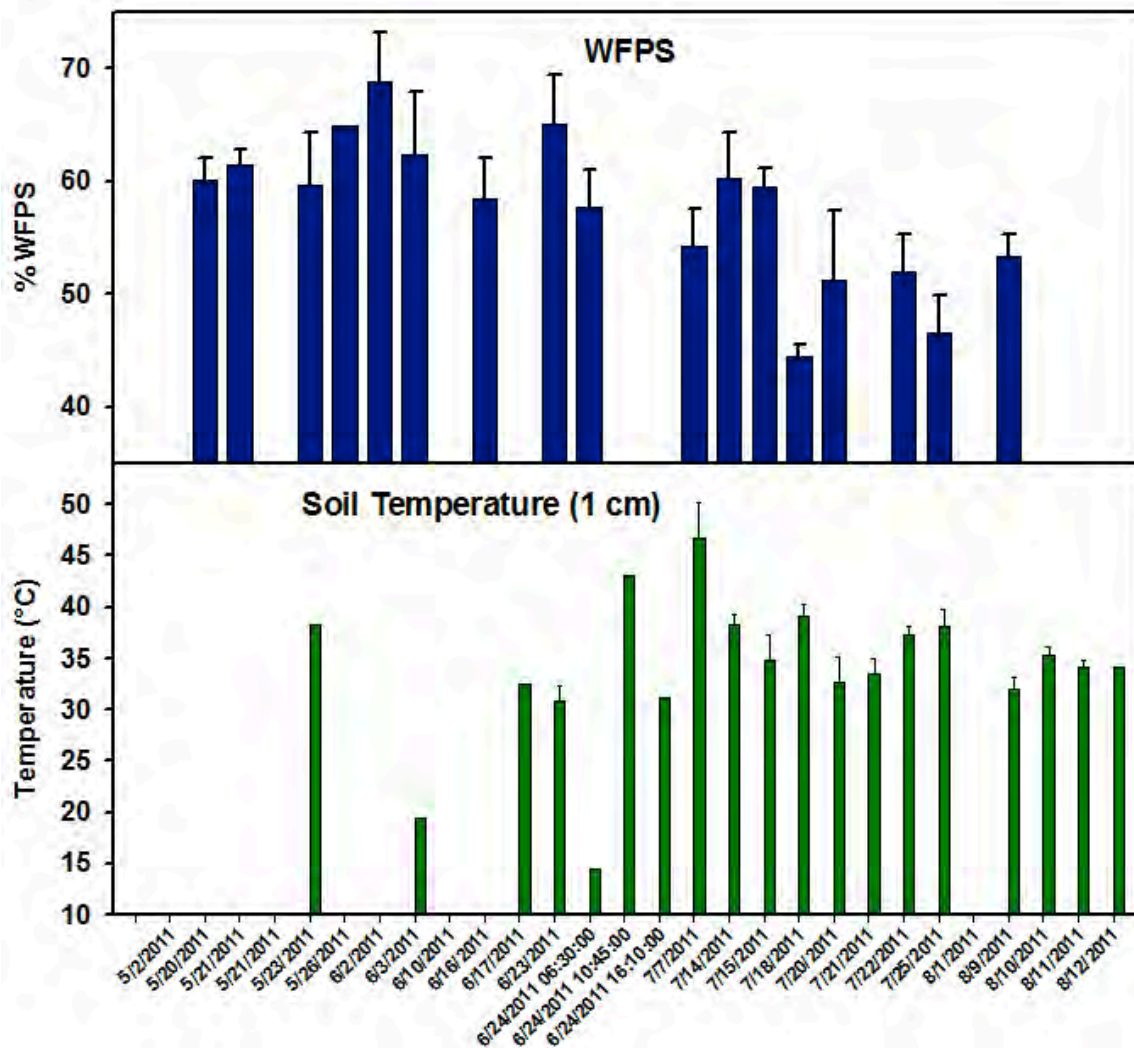




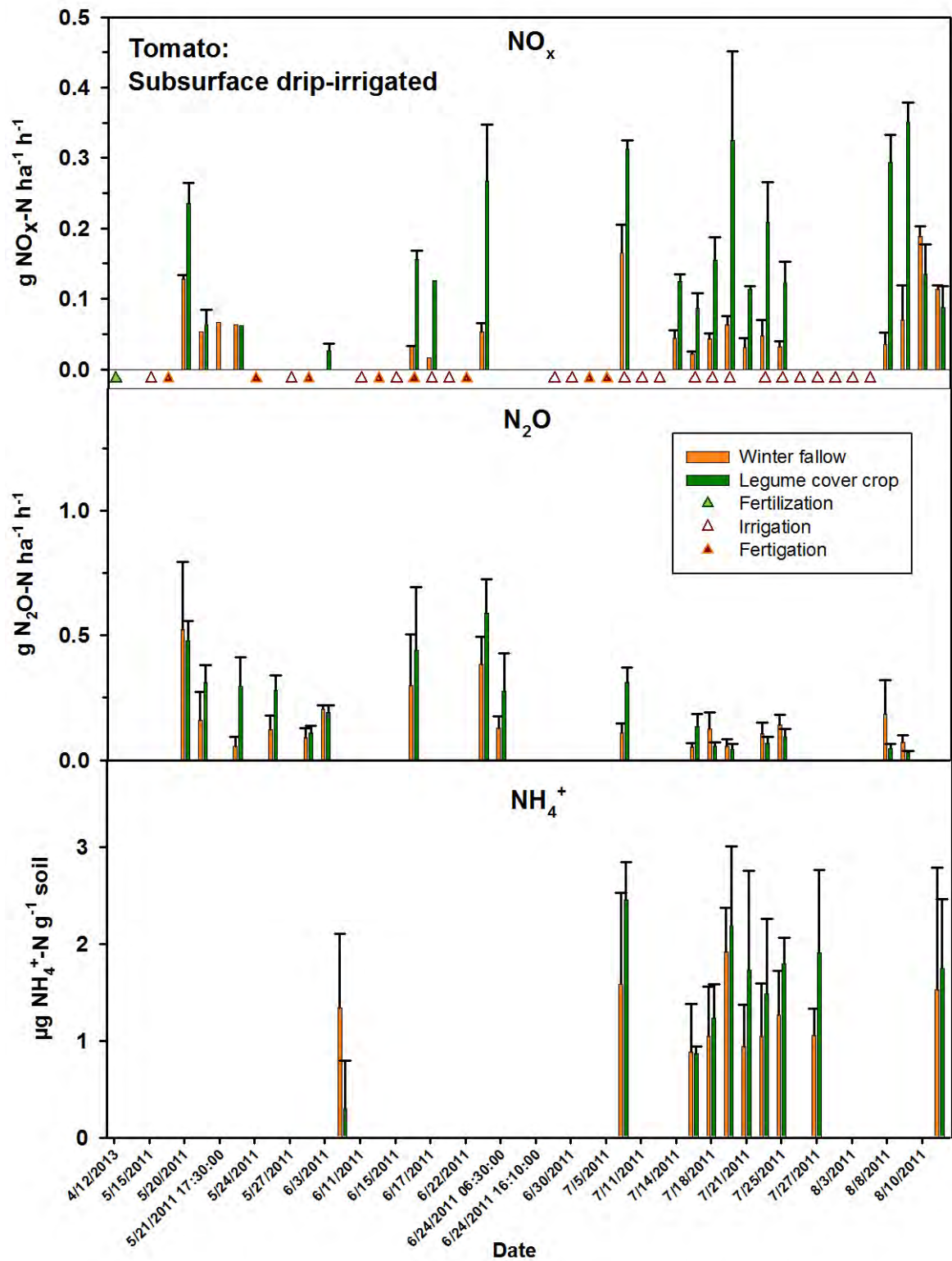
**Figure 4.** NO<sub>x</sub> and nitrous oxide (N<sub>2</sub>O) flux, soil temperature at 1 cm depth, water-filled pores space (WFPS), and soil ammonium (NH<sub>4</sub><sup>+</sup>) concentrations in furrow-irrigated tomato fertilized at 300, 162 and 0 kg N ha<sup>-1</sup>. Standard errors are shown as line bars. n=3. Triangles along the x-axis indicate inorganic fertilization (green) and irrigation (white) events.

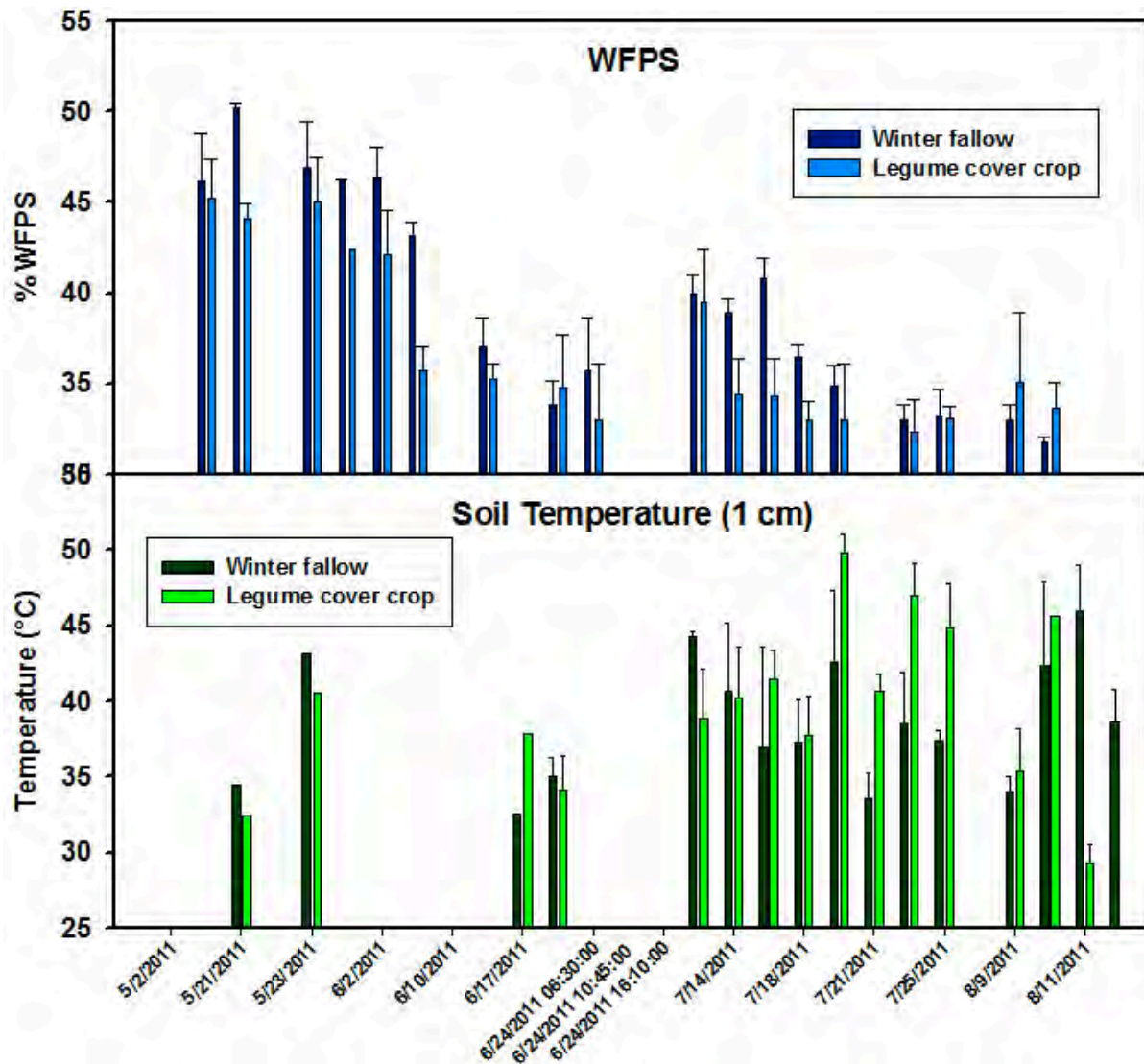






**Figure 5.** NO<sub>x</sub> and nitrous oxide (N<sub>2</sub>O) flux, soil ammonium (NH<sub>4</sub><sup>+</sup>) concentrations, water-filled pores space (WFPS), and soil temperature in the furrow-irrigated, winter cover-cropped tomato system fertilized with 179 kg N ha<sup>-1</sup> in cover cropped and winter-fallow tomato cropping systems. Standard errors are shown as line bars. n=3. Triangles along the x-axis indicate inorganic fertilization (green) and irrigation (white) events.





**Figure 6.**  $\text{NO}_x$  and nitrous oxide ( $\text{N}_2\text{O}$ ) flux, soil ammonium ( $\text{NH}_4^+$ ) concentrations, water-filled pores space (WFPS), and soil temperature in subsurface drip-irrigated tomato fertilized with  $179 \text{ kg N ha}^{-1}$  in cover cropped and winter-fallow tomato cropping systems. Standard errors are shown as line bars.  $n=3$ . Triangles along the x-axis indicate inorganic fertilizer application (green), irrigation (white) and fertigation (black) events.

### 3.4 Wheat

The  $\text{NO}_x$  fluxes measured in early summer in the wheat systems were between 9 and  $70 \text{ mg NO}_x\text{-N ha}^{-1} \text{ h}^{-1}$  (Table 8). The  $\text{N}_2\text{O-N}$  fluxes in the N fertilized treatments were 10 to 40 times greater than the  $\text{NO-N}$  fluxes, and the  $\text{NH}_4^+$  concentrations were between 1 and  $2.5 \mu\text{g NH}_4^+\text{-N g}^{-1}$  soil.

**Table 8.** The NO<sub>x</sub> and N<sub>2</sub>O flux, soil ammonium concentrations, and water filled pore space in five wheat treatments at Dixon and Russell Ranch field sites.

Date	NO <sub>x</sub> (mg NO-N ha <sup>-1</sup> h <sup>-1</sup> )	N <sub>2</sub> O (mg N <sub>2</sub> O-N ha <sup>-1</sup> h <sup>-1</sup> )	NH <sub>4</sub> <sup>+</sup> (μg NH <sub>4</sub> <sup>+</sup> -N g <sup>-1</sup> )	WFPS (%)
<b>May 24, 2011</b>				
Control	25.8 ± 7.7	10.0 ± 3.5	1.6 ± 1.2	46 ± 4
AA & U	17.8 ± 12.4	179.1 ± 60.5	1.9 ± 0.9	45 ± 4
AS & U	9 ± 10.9	108.5 ± 60.7	0.9 ± 0.2	46 ± 4
<b>May 30, 2012</b>				
Wheat-T-fallow	69.5 ± 22.7	1377 ± 242	1.4 ± 0.2	nd
Wheat-T-cc	55.4 ± 4.8	2487 ± 405	2.5 ± 0.4	nd

Control = no N fertilizer applied; AA & U = 112 kg N ha<sup>-1</sup> applied as anhydrous ammonia at planting and 98 kg N ha<sup>-1</sup> applied as urea in February; AS & U = 112 kg N ha<sup>-1</sup> applied as ammonium sulfate at planting and 98 kg N ha<sup>-1</sup> applied in February; Wheat-T-fallow = wheat-tomato-fallow rotation; wheat-T-cc = wheat-tomato-cover crop rotation, wheat was fertilized with 112 kg N applied as urea at planting and 80 kg N ha<sup>-1</sup> applied as foliar application in March.

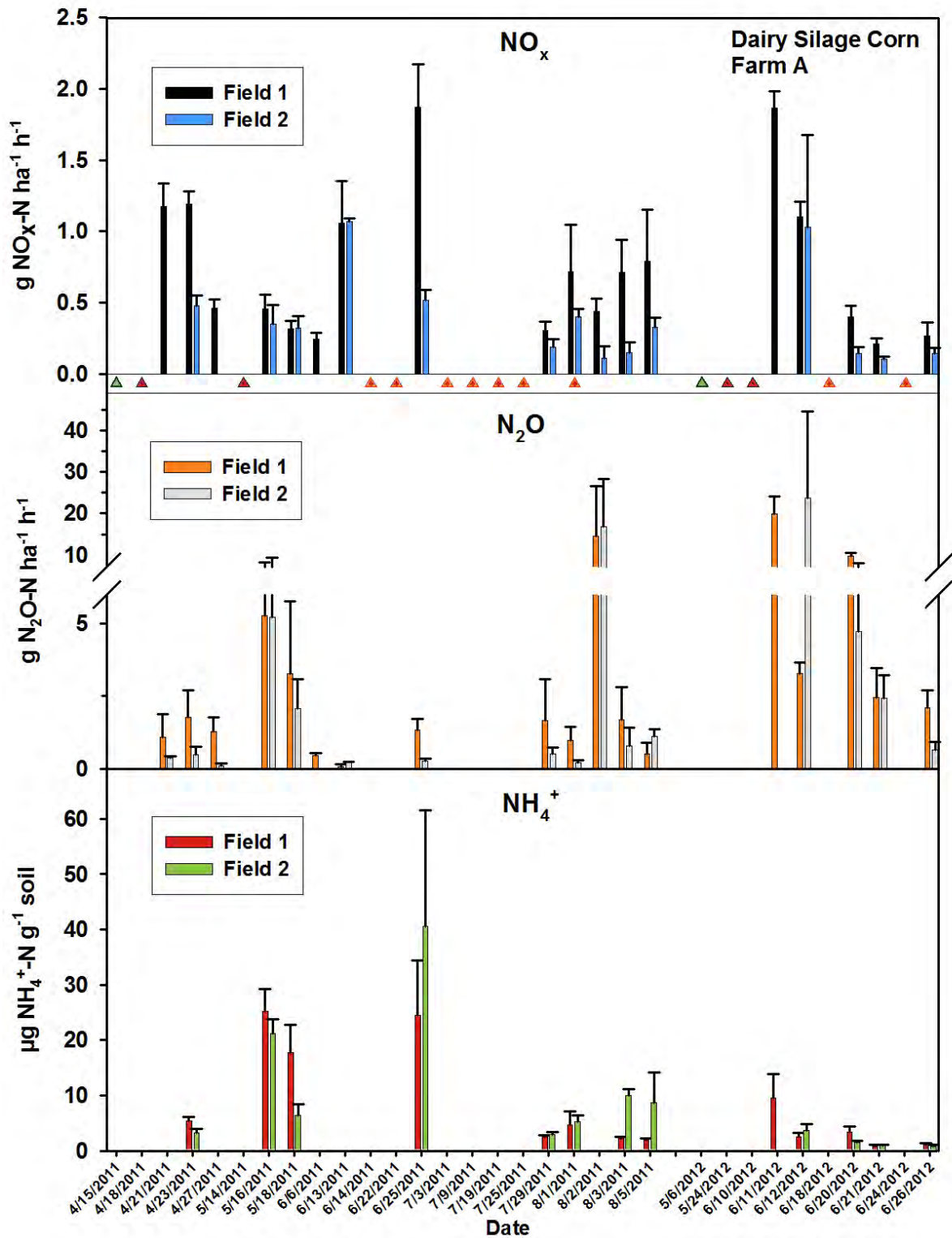
### 3.5 Dairy silage corn

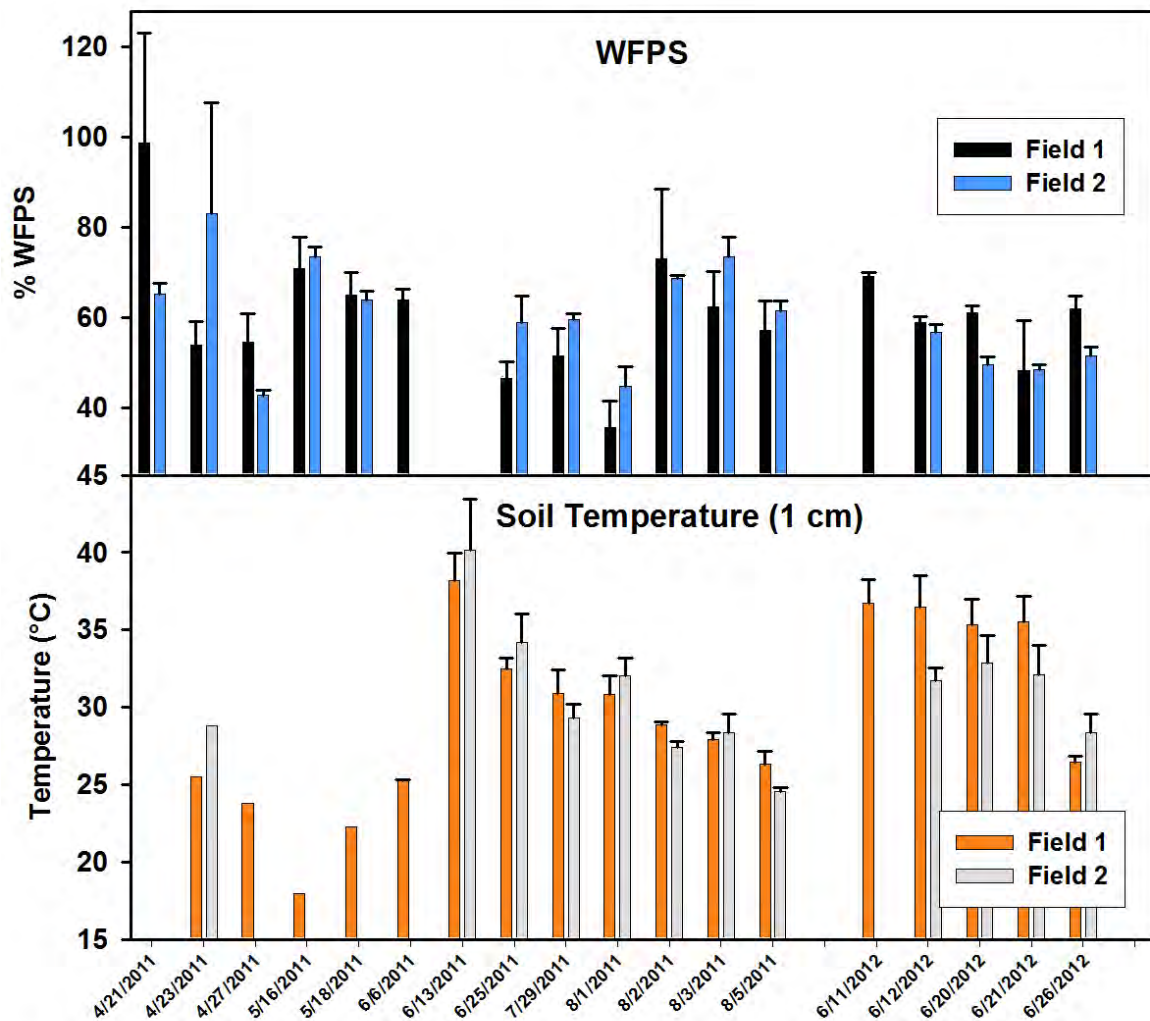
The soil temperatures (1 cm depth) during measurements at all the sites were 25-40°C (Figures 7, 9, 10). The WFPS ranged from 50-80% at Farm A and from 40-80% at Farm B, but on Farm C, all WFPS measurements were >70% and sometimes reaching 100%.

In the fields of Farm A, the NO<sub>x</sub> flux was mostly ≤1 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup>, except for two days when NO<sub>x</sub> flux reached about 2 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> (Figure 7). The soil NH<sub>4</sub><sup>+</sup> concentrations on one of those days was 25 μg NH<sub>4</sub><sup>+</sup>-N g<sup>-1</sup> (June 25, 2011), and 10 μg NH<sub>4</sub><sup>+</sup>-N g<sup>-1</sup> on the other (June 11, 2012). Soil NO<sub>2</sub><sup>-</sup> concentrations were low on most days, and there was no correlation of NO<sub>x</sub> flux with NO<sub>2</sub><sup>-</sup> concentration in the soil. Nevertheless, the highest NO<sub>x</sub> flux occurred when NO<sub>2</sub><sup>-</sup> was at the highest concentration (Figure 8). The N<sub>2</sub>O fluxes were mostly <5 g N<sub>2</sub>O-N ha<sup>-1</sup> h<sup>-1</sup>, reaching up to 20 g N<sub>2</sub>O-N ha<sup>-1</sup> h<sup>-1</sup> on some days.

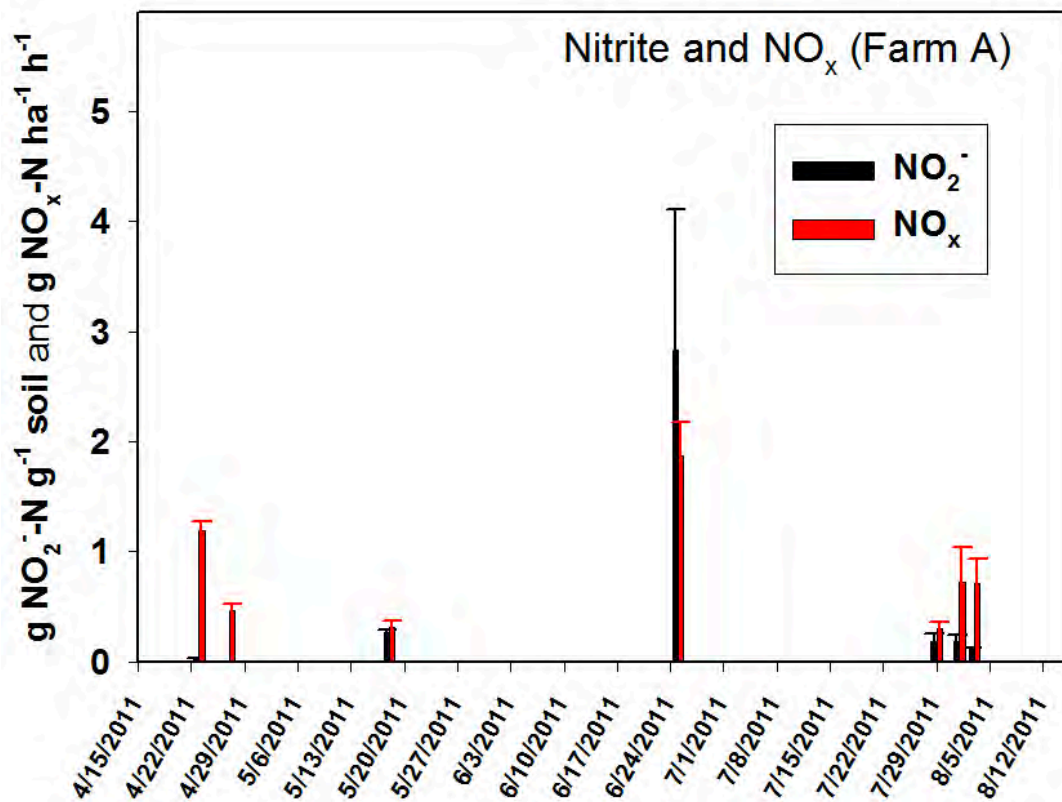
In two fields of Farm B, the NO<sub>x</sub> fluxes were mostly <2 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup>, but on two occasions (June 19, 2011 and June 15, 2012), NO<sub>x</sub> fluxes reached >14 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> (Figure 8). On those two occasions the soil NH<sub>4</sub><sup>+</sup> concentrations were 140 and 50 μg NH<sub>4</sub><sup>+</sup>-N g<sup>-1</sup> soil.

At Farm C, the NO<sub>x</sub> flux was mostly <1 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup>, except between June 22 and 27, 2011, when NO<sub>x</sub> fluxes reached up to 3.3 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> and between Sept 18 and 21, 2012, when NO<sub>x</sub> fluxes up to 41 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> were recorded (Figure 10). At Farm C, NO<sub>x</sub> fluxes in 2011 were mostly <0.5 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> except for three days (June 22 – June 27, 2011) when fluxes peaked at 3.3 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> (Figure 9). During that period, soil NH<sub>4</sub><sup>+</sup> concentrations were 7 μg NH<sub>4</sub><sup>+</sup>-N g<sup>-1</sup> and the WFPS was 96%. Soil NO<sub>2</sub><sup>-</sup> concentrations during this period were about 4 μg NO<sub>2</sub><sup>-</sup>-N g<sup>-1</sup> (Figure 11). In June 2012, the NO<sub>x</sub> flux exceeded 1 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> on only one day (June 27) when soil NH<sub>4</sub><sup>+</sup> concentrations were 24 μg NH<sub>4</sub><sup>+</sup>-N g<sup>-1</sup> and the WFPS was 76%. In September 2012, the NO<sub>x</sub> fluxes peaked at 42 g NO<sub>x</sub>-N ha<sup>-1</sup> d<sup>-1</sup> (September 21) when soil NH<sub>4</sub><sup>+</sup> concentrations were 8 μg NH<sub>4</sub><sup>+</sup>-N g<sup>-1</sup> and the WFPS was 88%. The N<sub>2</sub>O flux at Farm C in both years was elevated in June, with fluxes up to 30-40 g N<sub>2</sub>O-N ha<sup>-1</sup> h<sup>-1</sup>, but in September 2012, peak N<sub>2</sub>O fluxes were lower than the NO<sub>x</sub> fluxes.



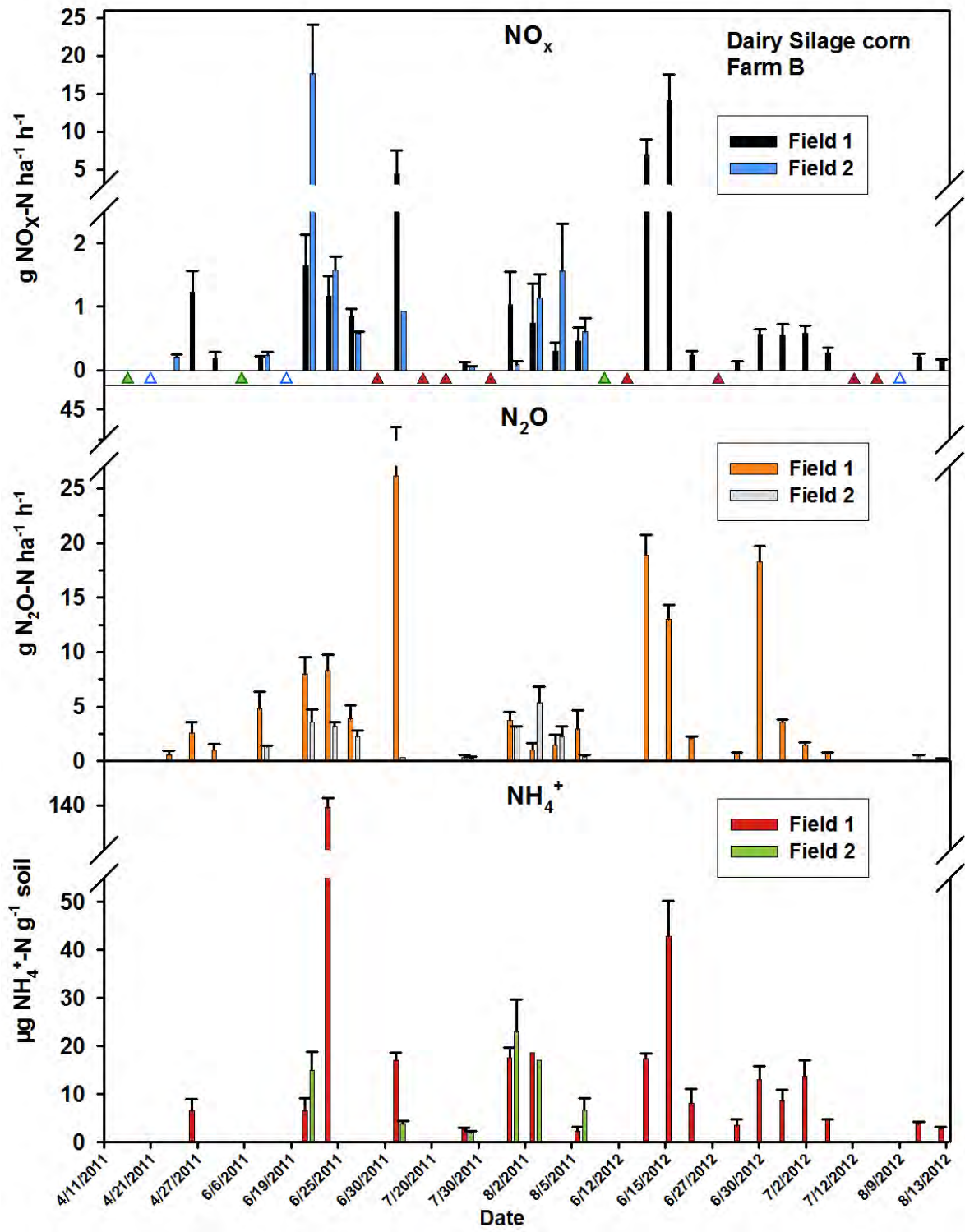


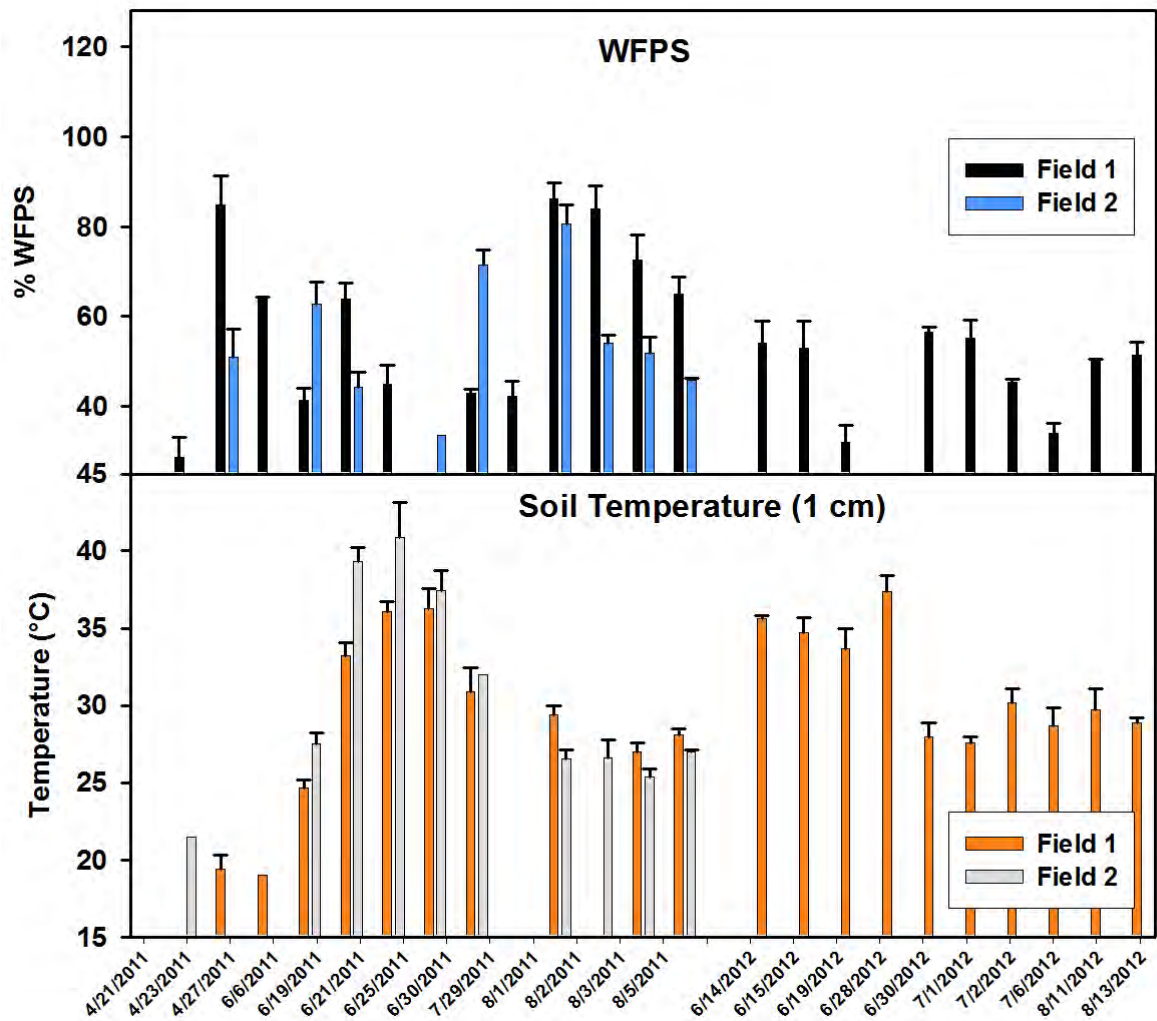
**Figure 7.** NO<sub>x</sub> and nitrous oxide (N<sub>2</sub>O) flux, soil ammonium (NH<sub>4</sub><sup>+</sup>) concentrations, water-filled pores space (WFPS), and soil temperature in the dairy silage corn systems of Farm A. Standard errors are shown as line bars. n=4. Triangles along the x-axis indicate inorganic fertilization (green), irrigation with lagoon water (red), and irrigation with lagoon water and inorganic fertilizers (orange) events.



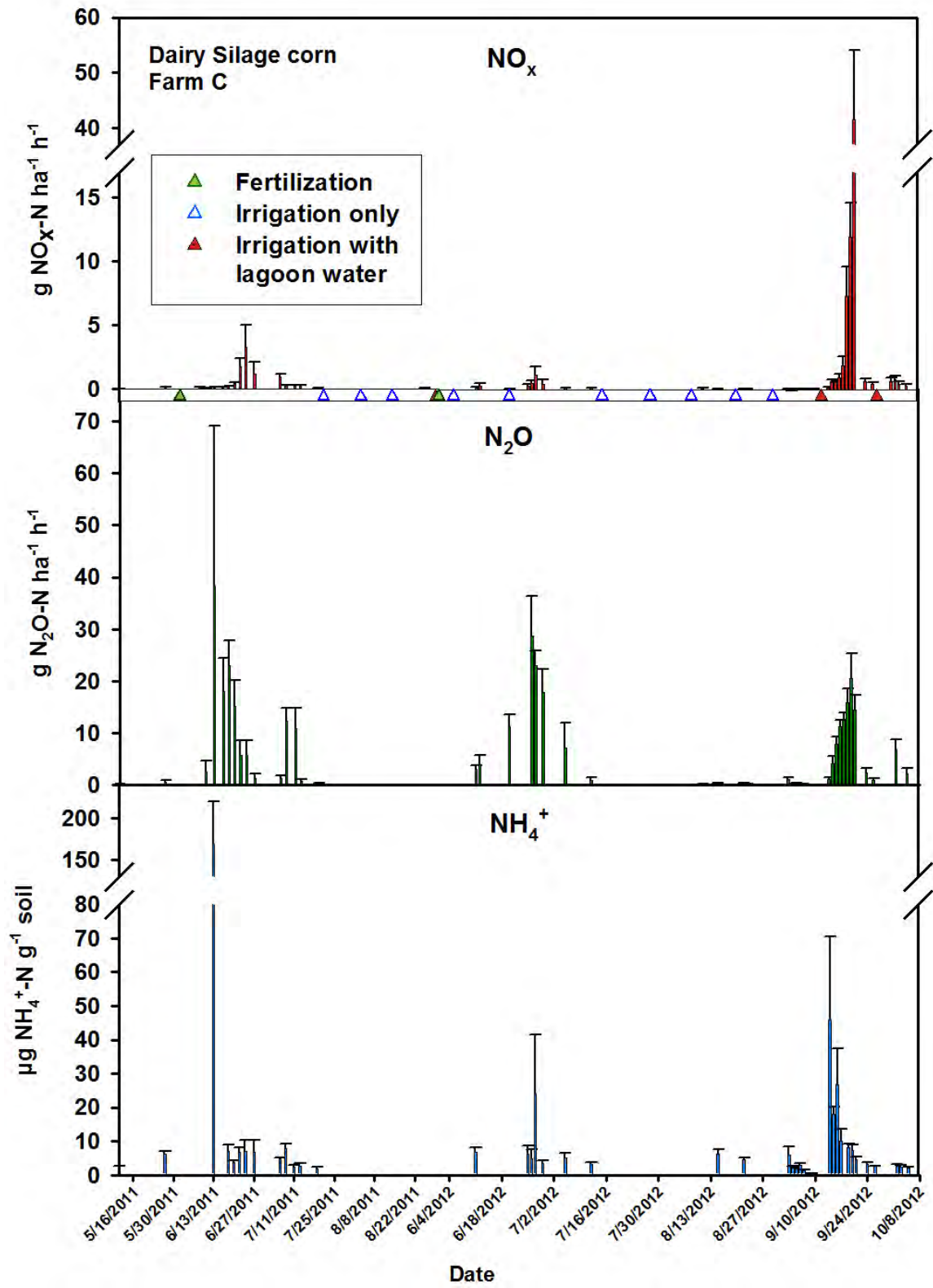
**Figure 8.** Soil nitrite concentrations and NO<sub>x</sub> flux at Farm A. Standard errors shown as line bars. n=4.

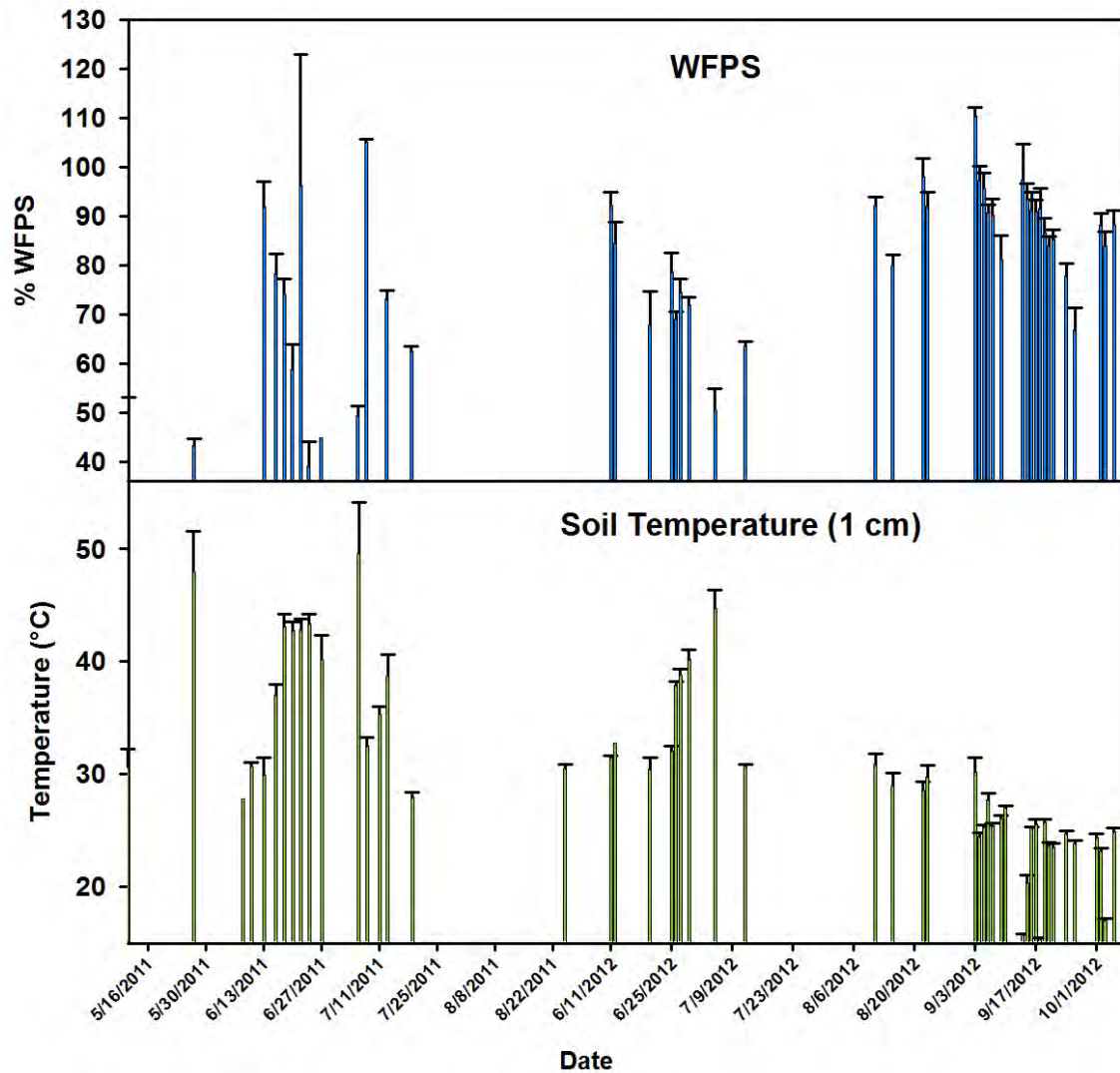




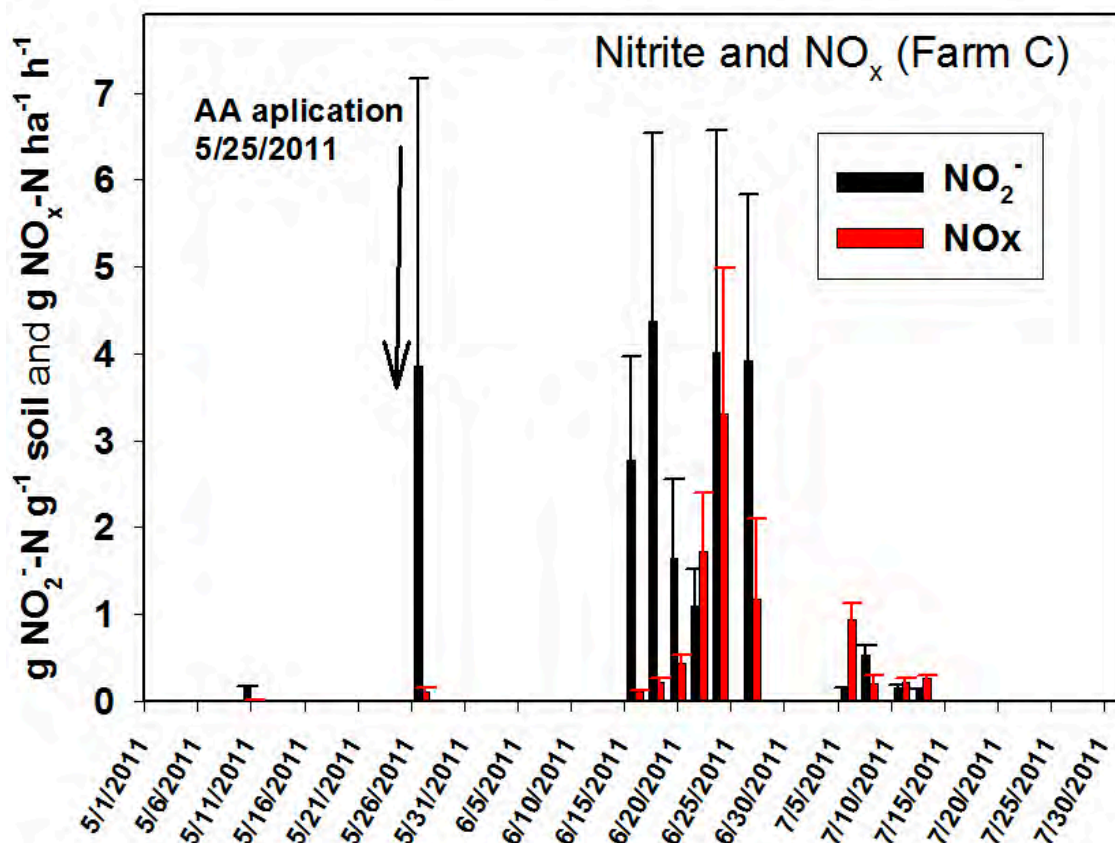


**Figure 9.** NO<sub>x</sub> and nitrous oxide (N<sub>2</sub>O) flux, soil ammonium (NH<sub>4</sub><sup>+</sup>) concentrations, water-filled pore space (WFPS), and soil temperature in the dairy silage corn systems of Farm B. Standard errors are shown as line bars. n=4. Triangles along the x-axis indicate inorganic fertilization (green), irrigation (white), and irrigation with lagoon water (red) events.





**Figure 10.** NO<sub>x</sub> and nitrous oxide (N<sub>2</sub>O) flux, soil ammonium (NH<sub>4</sub><sup>+</sup>) concentrations, water-filled pore space (WFPS), and soil temperature in the dairy silage corn systems of Farm C. Standard errors shown as line bars. n=4. Triangles along the x-axis indicate inorganic fertilization (green), irrigation (white), and irrigation with lagoon water (red) events.



**Figure 11.** Soil nitrite concentrations vs. NO<sub>x</sub> flux at Farm C. The date of the anhydrous ammonia (AA) application is also shown. Standard errors shown as line bars. n=4.

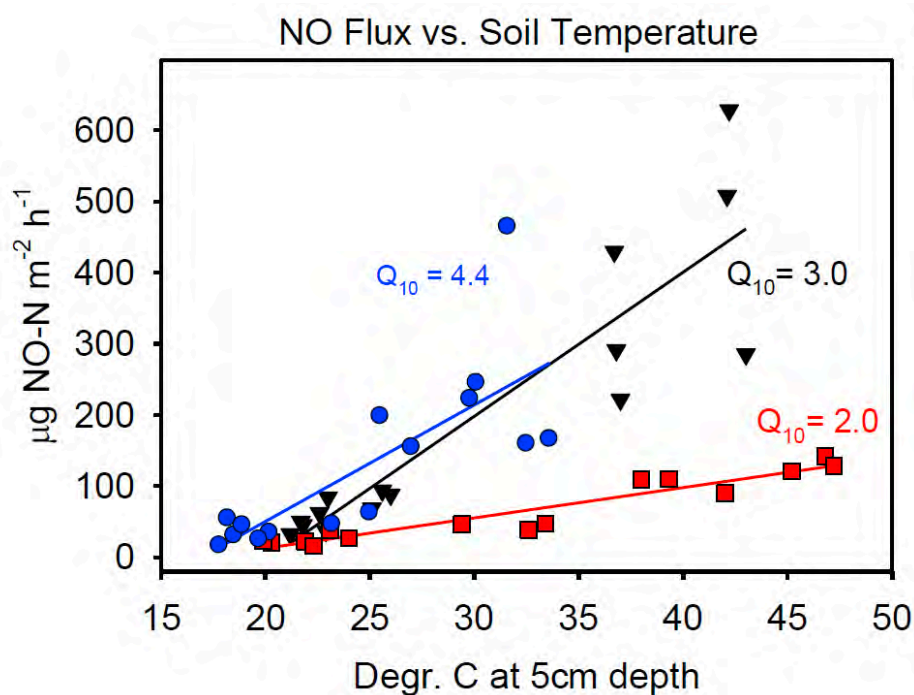
### 3.6 Relationship between NO<sub>x</sub> flux and soil temperature

The NO<sub>x</sub>-flux in response to temperature change, expressed as the change in rate per 10°C change in soil temperature (Q<sub>10</sub>) at 1 and 5 cm depth, was determined in 8 experiments at some of the field sites. The Q<sub>10</sub> ranged from 1.2-3.8 with a mean of 2.5 based on soil temperature changes at 1 cm depth, and from 1.3-5.2 with a mean of 3.4 based on soil temperature changes at 5 cm depth (Table 9). Figure 12 shows data from three sites adjacent to and inside a corn field. Each data point shown represents an individual measurement.

**Table 9.** Q<sub>10</sub> of NO<sub>x</sub>-flux based on the change in soil temperature at 1 and 5 cm depths.

Location	Date	Min. T. 1 cm (°C)	Max. T. 1 cm (°C)	Min. T. 5 cm (°C)	Max. T. 5 cm (°C)	Q <sub>10</sub> 1 cm	Q <sub>10</sub> 5 cm
C1	7/21/12	18.8	45.8	21.2	42.2	1.8	2.0
C2	7/21/12	18.2	49.9	21.7	46.8	2.4	3.0
C3	7/21/12	17.1	32.5	18.2	32.5	3.8	4.4
FB	8/10/12	19.8	29.2	20.4	27.6	1.6	2.1
FC	9/15/12	15.6	25.3	17.5	23.5	3.0	4.2
FC	9/16/12	15.3	25.0	16.0	23.2	1.9	2.5
FC	9/26/12	16.8	24.9	16.9	23.0	1.2	1.3
FC	10/3/12	15.9	25.2	17.2	23.6	3.0	5.2
Median						2.2	2.8
Mean						2.5	3.4

C1-C3 = Campbell Tract, UC Davis; FB = Farm B; FC = Farm C



**Figure 12.**  $\text{NO}_x$ -flux in relation to soil temperature at 5 cm during the course of one day at three locations. The fluxes represented by the red and black symbols were measured at the edge of a corn field (WFPS 43%), the fluxes represented by the blue symbols were measured within the same corn field (WFPS 46%).

#### 4. Discussion

We measured  $\text{NO}_x$  fluxes under varied soil moisture conditions in five different cropping systems receiving N inputs at different rates and in varied forms, and we calculated average hourly fluxes for each treatment and system. The average hourly fluxes provide information on the general trends in emissions among the different systems studied (Table 7). The lowest fluxes were observed in the wheat systems, the tractor rows of the almond orchard, the furrow-irrigated tomato control treatment (no N fertilizer applied this season) and the two SDI treatments of the tomato systems (Table 7). Somewhat greater average hourly  $\text{NO}_x$  fluxes were measured in alfalfa, the tree rows of the almond orchard, and the FI treatments of the tomato systems fertilized at recommended N rates. Our results are in agreement with those of earlier studies in California cropping systems. For alfalfa, except for one day, the measured fluxes in this study ranged from 0.07-0.6  $\text{g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$ , which is very similar to earlier reported ranges of 0.1-0.6 (Matson and Firestone, 1995). Likewise, the range of emissions in the sprinkler irrigated almond orchard in our study was 0.02-1.09 vs. 0.1-0.9  $\text{g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$  in the previous study, and for FI tomatoes the corresponding ranges are 0.04-1.2 vs. 0.1-1.2  $\text{g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$  (Matson and Firestone, 1995). In the systems receiving high N inputs, such as FI tomatoes fertilized with 300  $\text{kg N ha}^{-1}$  and the dairy silage corn systems, the average  $\text{NO}_x$  fluxes were between 0.4 and 2.8  $\text{g NO}_x\text{-N ha}^{-1} \text{h}^{-1}$  and higher than in the other systems studied here. There are not many field data of  $\text{NO}_x$  emissions resulting from liquid manure applications in the literature. Although on most days the range of fluxes measured by us in corn systems was similar as in a previous study (0.01-5.2  $\text{g NO}_x\text{-N ha}^{-1}$

h<sup>-1</sup>) (Matson and Firestone, 1995), we also recorded some fluxes that exceeded those values by a wide margin and were comparable to those reported for fertilized (184 kg N as urea and 63 kg N as anhydrous ammonia) irrigated wheat systems in Sonora, Mexico (peak fluxes of 20-55 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup>) (Matson *et al.*, 1998), and with the fluxes measured above an injection band of anhydrous ammonia (120 kg N ha<sup>-1</sup>) in a California tomato system (5-10 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup>, with peak values of up to 100 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup>) (Venterea and Rolston, 2000b).

Nitric oxide flux is highly dependent on temperature, with a predicted doubling of NO production for each 10°C increase in temperature between 15° and 35°C (Williams *et al.*, 1995). Most of our NO<sub>x</sub> flux measurements were made in the afternoons. The recorded fluxes therefore represent values close to the maximum of those days. The fluxes, soil temperatures, and times of measurements are compiled in the Appendix (Tables 1A-6A). If the reported NO<sub>x</sub> emission values will be used as modeling inputs, the fluxes need to be adjusted for diel fluctuations in temperature. We derived Q<sub>10</sub> factors under field conditions in three different corn cropping systems. The lowest NO<sub>x</sub> flux of the day was always measured early in the morning when soil temperatures typically are at their daily minimum. The ranges of Q<sub>10</sub> varied from 1.2 to 3.8 with a mean of 2.5 and a median value of 2.2 based on soil temperature dynamics at a depth of 1 cm. Based on soil temperature at 5 cm, the range was 1.3-5.2 with a mean of 3.5 and a median of 2.8 (Table 15). The Q<sub>10</sub> obtained based on soil temperature at 1 cm may have been lower because nitrifying bacteria have been reported to stop growing >40°C (Focht and Verstraete, 1977). Temperatures at 1 cm depth were approaching or exceeding this temperature on some of the days and were higher than at 5 cm depth.

Besides temperature, availability of the substrate NH<sub>4</sub><sup>+</sup> and NO<sub>2</sub><sup>-</sup>, as influenced by the amount, form, and placement of N inputs, controls NO<sub>x</sub> flux. For example, on Farm B, NO<sub>x</sub> fluxes of 10-15 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> were measured shortly after UAN32 fertilizer applications on June 15, 2011 and June 7, 2012, and an irrigation with lagoon water. The high NO<sub>x</sub> emissions also coincided with NH<sub>4</sub><sup>+</sup> concentrations >100 µg NH<sub>4</sub><sup>+</sup>-N g<sup>-1</sup>. In the tomato system, the application of 300 kg N ha<sup>-1</sup> as UAN32 – almost twice the recommended N rate for furrow-irrigated tomato crops – lead to sustained high NO<sub>x</sub> emissions lasting about 6 weeks. It is possible that in the dairy silage corn systems the liquid manure enhanced NO<sub>x</sub> emissions due to the availability of carbon which stimulates O<sub>2</sub> consumption in the soil. Nitric oxide and N<sub>2</sub>O production increase with decreasing O<sub>2</sub> levels in the soil (Zhu *et al.*, 2013). Nitrite, the intermediate during ammonia oxidation to NO<sub>3</sub><sup>-</sup>, is a precursor of NO and N<sub>2</sub>O in the soil (VanCleemput and Samater, 1996; Venterea and Rolston, 2000a, b; Zhu *et al.*, 2013). We detected NO<sub>2</sub><sup>-</sup> after application of anhydrous ammonia (Farm C) and after application of liquid manure and UAN in the irrigation water at Farm A, and in both cases enhanced NO<sub>x</sub> emissions coincided with high NO<sub>2</sub><sup>-</sup> levels in the soil (Figures 8 and 11). There was also a weak correlation of NO<sub>x</sub> flux with NO<sub>2</sub><sup>-</sup> levels in the almond orchard (Figure 2).

Since most NO produced in soil is consumed to produce N<sub>2</sub>O (Venterea and Rolston, 2000b; Venterea *et al.*, 2004), the placement of the substrate can be expected to affect the magnitude NO<sub>x</sub> emissions. Diffusion of gases at high WFPS is low, and consumptive processes of NO are likely favored over the release of this reactive gas to the atmosphere. In general, during the periods when high NO<sub>x</sub> emissions occurred, the WFPS was ≤60%, but there were some noteworthy exceptions. For example, in September 2012, NO<sub>x</sub> fluxes on the order of 7-42 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup> occurred at Farm C while WFPS was >90%. The high NO<sub>x</sub> emissions took place in spite of high WFPS most likely because the liquid manure, which was applied via flood irrigation, provided the substrate (NH<sub>4</sub><sup>+</sup>) for nitrification and NO<sub>x</sub>

production near the soil surface. It is interesting that in June, 2011, substantial  $\text{NO}_x$  flux following the anhydrous ammonia application at Farm C did not occur until WFPS declined to <50% (Figure 10) even though  $\text{NO}_2^-$  concentrations were elevated as soon as the anhydrous ammonia was applied (Figure 11).

The  $\text{NO}_x$  emissions were related to N inputs and  $\text{NH}_4^+$  availability, and most  $\text{NO}_x$  emitted was likely due to nitrification. Once most of the applied  $\text{NH}_4^+$  was converted to  $\text{NO}_3^-$  or taken up by the crops, the  $\text{NO}_x$  fluxes subsided. This was particularly evident at Farm C, where lower  $\text{NH}_4^+$  concentrations in the soil coincided with tapering off of  $\text{NO}_x$  flux. Similarly, as soil  $\text{NH}_4^+$  concentrations decreased from  $>100 \mu\text{g NH}_4^+\text{-N g}^{-1}$  on Farm B, the  $\text{NO}_x$  flux subsided to more moderate levels. In the almond orchard, following four fertigation events, there were four distinct peaks of  $\text{NO}_x$  emissions, which declined over the course of 10 days probably because the  $\text{NH}_4^+$  was taken up by the trees or nitrified although the decrease in  $\text{NH}_4^+$  concentrations was not clearly shown. It appears, therefore, that in general  $\text{NO}_x$  fluxes decrease with time since N fertilization, which has been observed earlier (Williams *et al.*, 1992; Matson and Firestone, 1995).

The relationship between  $\text{N}_2\text{O}$  and  $\text{NO}_x$  fluxes varied, depending on the system. Average hourly  $\text{NO}_x$  fluxes were similar (SDI-irrigated and wheat systems, as well as Field 2 on Farm B) or lower than average hourly  $\text{N}_2\text{O}$  fluxes, except for furrow-irrigated tomatoes fertilized with an excessive N rate, which had higher average  $\text{NO}_x$  than  $\text{N}_2\text{O}$  fluxes (Table 7). Differences between  $\text{NO}_x$  and  $\text{N}_2\text{O}$  fluxes in the different systems may be explained by soil water content and ultimately by irrigation systems. While  $\text{N}_2\text{O}$  production can be expected to be related to  $\text{NO}_x$  production, the opportunity for the consumption of  $\text{NO}_x$  is greater at high than low and intermediate soil water content. Therefore, when WFPS is high,  $\text{N}_2\text{O}$  tend to be greater than  $\text{NO}_x$  emissions. The flood-irrigated alfalfa and silage corn systems, where average  $\text{N}_2\text{O}$  fluxes were 2 to 16 times higher than average  $\text{NO}_x$  fluxes in six of the seven fields, reached high WFPS with every irrigation, and this likely explains the larger differences between  $\text{N}_2\text{O}$  and  $\text{NO}_x$  fluxes in the flood- than drip- and furrow-irrigated systems.

To compare the contribution of  $\text{NO}_x$  from agricultural soil to overall statewide  $\text{NO}_x$  emissions, the average values measured in the present study were used to calculate a rough estimate of total  $\text{NO}_x$  emissions from agricultural soil during summer. There are many caveats to deriving such an estimate. First, the  $\text{NO}_x$  fluxes were measured in only 16 fields in five major cropping systems whereas  $>300$  different crops are grown in California. Notably absent in the present study were measurements in grapevine, rice, citrus, cotton, pasture land, and others. Second, the measured  $\text{NO}_x$  fluxes were highly variable, generally decreasing with time since fertilizer applications, and the average hourly  $\text{NO}_x$  fluxes of this study are not necessarily representative for certain days when the actual fluxes could potentially be much higher or lower than the average fluxes. Third, site-specific differences due to management practices and soil types were not considered when extrapolating the results of the present study to other cropping systems. The approximate estimates of  $\text{NO}_x$  emissions (in  $\text{Mg NO}_x \text{ d}^{-1}$ , or tonnes  $\text{NO}_x$  per day) (Table 10) were derived in two ways. Either the hourly  $\text{NO}_x$  fluxes measured in this study were assumed to represent the daytime fluxes (12 hours) and the night-time fluxes (12 hours) were assumed to be 50% of the daytime fluxes ('low' estimate), or the hourly fluxes were considered representative for the total daily (24 hours) emissions ('high' estimate). The average  $\text{NO}_x$  fluxes in furrow-irrigated tomato were assumed to be representative for all vegetable, field & seed, and rice cropping systems, whereas all fluxes in fruit & nut cropping systems were assumed to be similar as the ones measured in the almond orchard. Average fluxes in alfalfa were assumed to be representative for all alfalfa and forage



crops, and those in corn systems of the dairy farms were considered representative for all silage corn acreage.

**Table 10.** Estimates of daily NO<sub>x</sub> emissions from agricultural land in California.

Cropping systems	Area (ha) <sup>†</sup>	Hourly NO <sub>x</sub> flux*	Daily NO <sub>x</sub> flux (low est.)	Daily NO <sub>x</sub> flux (high est.)
		g NO <sub>x</sub> -N ha <sup>-1</sup> h <sup>-1</sup>	Mg NO <sub>x</sub> d <sup>-1</sup>	Mg NO <sub>x</sub> d <sup>-1</sup>
Vegetables	517,004	0.32 <sup>1</sup>	9.8	13.0
Fruits & nuts	989,069	0.35 <sup>2</sup>	20.5	27.3
Field & seed crops	332,794	0.32 <sup>1</sup>	6.3	8.4
Rice	234,818	0.32 <sup>1</sup>	4.4	5.9
Wheat	319,838	0.04 <sup>3</sup>	0.8	1.0
Alfalfa & forage	724,696	0.54 <sup>4</sup>	23.1	30.9
Silage corn	192,308	1.35 <sup>5</sup>	15.3	20.5
Total	8,177,000		80.3	107.0

<sup>†</sup>Based on California Department of Food & Agriculture 2011 data, \* estimated from hourly NO<sub>x</sub> flux in <sup>1</sup>furrow-irrigated tomato fertilized with 162 kg N ha<sup>-1</sup>, <sup>2</sup>microjet sprinkler-irrigated almond, <sup>3</sup>wheat, <sup>4</sup>flood-irrigated alfalfa, and <sup>5</sup>flood-irrigated corn fertilized with mineral fertilizer and liquid dairy manure.

The total NO<sub>x</sub> emissions in California have declined from about 4,900 Mg d<sup>-1</sup> in 1990 to 3,000 Mg d<sup>-1</sup> in 2010 and are projected to decrease to 2,200 Mg d<sup>-1</sup> by 2020 (CARB). Therefore, based on the above estimate, NO<sub>x</sub> emissions from agricultural soil account for <4% of total NO<sub>x</sub> emitted, but the contribution of NO<sub>x</sub> from agricultural land could increase to about 5% within this decade if further reductions of NO<sub>x</sub> from mobile and stationary sources will be attained.

## Summary and Conclusions

Emissions of NO<sub>x</sub> were measured in almond, alfalfa, tomato, wheat, and silage corn cropping systems during summer months (June-September) in California's Central Valley. The study was undertaken to estimate the contribution of NO<sub>x</sub> from agricultural soil in order to improve the predictive power of ozone (O<sub>3</sub>) models for the San Joaquin Valley because NO<sub>x</sub> availability under certain circumstances (e.g. when the availability of volatile organic compounds is high) controls O<sub>3</sub> formation. The NO<sub>x</sub> fluxes were measured in 17 different fields or treatments representing varied soil moisture conditions, nitrogen availability, and management practices, such as irrigation and nitrogen inputs.

The average NO<sub>x</sub> fluxes were lowest in wheat, the non-irrigated sections of an almond orchard and in subsurface drip-irrigated tomato (average flux <0.1 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup>), and intermediate in alfalfa, the irrigated sector of an almond orchard, and furrow-irrigated tomato N fertilized at recommended rates (average flux ≤0.5 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup>). The highest NO<sub>x</sub> emissions were measured in furrow-irrigated tomato fertilized at an excessive N rate and in silage corn systems receiving high N inputs in the form of synthetic N fertilizers and manure from dairy farms (average flux up to 3 g NO<sub>x</sub>-N ha<sup>-1</sup> h<sup>-1</sup>). The ranges of emissions near their daily maximum were comparable to those measured in earlier studies in the different systems, but following high N inputs, the emissions were higher (by an order of magnitude) for short periods (days).

The emissions at each location varied over time, depending on soil moisture, time since N fertilization, and soil temperature. In the present study, field experiments showed that NO<sub>x</sub> emissions increase on average 2.5- and 3.5-fold with each increase of 10°C in soil temperature at 1 and 5 cm depth, respectively. Enhanced NO<sub>x</sub> fluxes occurred under intermediate soil water contents (water-filled pore space 30-60%), whereas in relatively dry soils or at high water content, NO<sub>x</sub>-fluxes were low. The results suggest that NO<sub>x</sub> emissions are related to ammonium availability and nitrification rates, hence the decline of NO<sub>x</sub> flux with time since N fertilizer applications.

Based on this relatively limited data set given the great variety of cropping systems in the San Joaquin Valley, it appears that N fertilization at recommended N rates does lead to fairly predictable NO<sub>x</sub> emissions. However, the magnitude and duration of enhanced NO<sub>x</sub> emissions (increased by an order of magnitude or more), are not necessarily predictable because they are event-based (e.g. date of N fertilization) and depend on complex interactions among NO production, gas transport and NO consumption in the soil, as well as other variables such as soil temperature at different depths.

## **Recommendations**

The objective of this study was to provide field estimates of NO<sub>x</sub> emissions from agricultural land. Because NO<sub>x</sub> flux varies depending on farm management events (e.g. N fertilization, irrigation), day-by-day, or hourly, regional quantification of NO<sub>x</sub> emissions from agricultural sources would require elaborate models including the mosaic of cropping systems and their associated management. The average NO<sub>x</sub> emissions or ranges of emissions generated in this study might be extrapolated according to crop acreages and compared to known estimates of NO<sub>x</sub> production from mobile and other NO<sub>x</sub> sources as a rough estimate assessing the relative importance of NO<sub>x</sub> emissions from agricultural land under current conditions.

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## Glossary of Terms, Abbreviations and Symbols

AB32	Assembly Bill 32
banded	Concentrated line of material in soil, such as fertilizer applied by injection into the soil
CARB	California Air Resources Board
C	Carbon
°C	Degree(s) Celsius
cc	Cover crop
CrO <sub>3</sub>	Chromium trioxide
est.	estimated
Fertigation	Application of fertilizer dissolved in irrigation water
FI	Furrow irrigation
h	Hour(s)
ha	Hectare
H <sub>2</sub> O	Water
N	Nitrogen
NH <sub>4</sub> <sup>+</sup>	Ammonium
NO	Nitric oxide
NO <sub>2</sub> <sup>-</sup>	Nitrite
NO <sub>3</sub> <sup>-</sup>	Nitrate
NO <sub>x</sub>	Nitric oxide and nitrogen dioxide
N <sub>2</sub> O	Nitrous oxide
O <sub>2</sub>	Oxygen
O <sub>3</sub>	Ozone
PTFE	Teflon
PVC	Poly-vinyl chloride
SDI	Subsurface drip irrigation
SE	Standard error
UAN32	Urea ammonium-nitrate
UC	University of California
VOC	Volatile organic compounds
WFPS	Water-filled pore space

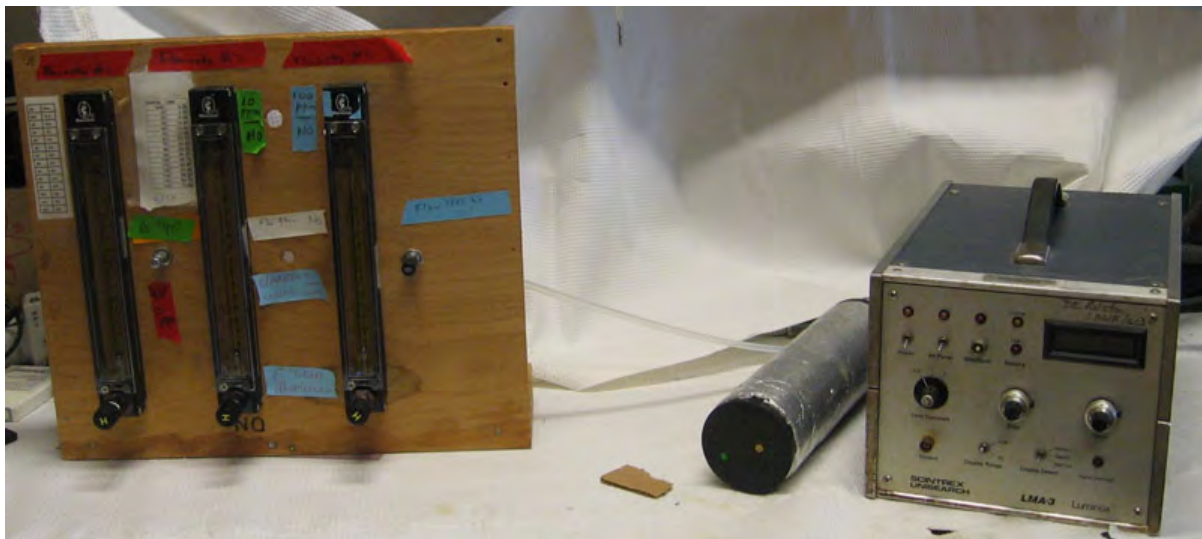
yr

Year

## APPENDIX



**Figure 1A.** Paraphernalia to measure NO<sub>x</sub> flux in the field. From left, vessel containing drying agent and Nafion™ tubing to remove air humidity, battery-powered Scintrex (LMA-3) NO<sub>x</sub> analyzer, chamber used in alfalfa fields, chamber used in corn, tomato, and wheat systems.



**Figure 2A.** Floating ball flowmeters (Manostat) used to measure the flow rates of NO standard gas and NO<sub>x</sub>-free air during calibration of the NO<sub>x</sub> analyzer.